REVIEW

Open Access

Damage mechanism and therapy progress of the blood-brain barrier after ischemic stroke

Hui-min Gao¹, Hao Chen², Gui-Yun Cui^{1,2} and Jin-Xia Hu^{1,2,3*}



Abstract

The blood-brain barrier (BBB) serves as a defensive line protecting the central nervous system, while also maintaining micro-environment homeostasis and inhibiting harmful materials from the peripheral blood. However, the BBB's unique physiological functions and properties make drug delivery challenging for patients with central nervous system diseases. In this article, we briefly describe the cell structure basis and mechanism of action of the BBB, as well as related functional proteins involved. Additionally, we discuss the various mechanisms of BBB damage following the onset of an ischemic stroke, and lastly, we mention several therapeutic strategies accounting for impairment mechanisms. We hope to provide innovative ideas for drug delivery research via the BBB.

Keywords Ischemic stroke, Blood-brain barrier, Stroke mechanism

Introduction

Paul Ehrlich [1] made the initial discovery that the dyestuff, when injected into the blood vessels, did not color brain parenchyma. Later, his student Edwin Goldmann [2] made additional observations that showed that the same dyestuff would stain brain tissue if injected into the cerebrospinal fluid. This led to the development of the vague concept of biological barriers between the blood and the brain parenchyma [3]. After conducting extensive research, Dr. Lena Stern presented the term "Barrière hématoencéphalique" to the faculty of Medicine in Geneva [4]. Furthermore, she has published a conceptual article on the topic of the BBB. Following the concrete conceptualization of the BBB, which was developed by Paul Ehrlich, Edwin Goldmann, and Lena Stern, extensive and ongoing studies have been conducted on its morphology, molecular composition, and physiological properties [5]. The BBB maintains the steady-state of the neural system and is therefore considered to be an important structure. Numerous studies have investigated the function of the BBB. Ischemic stroke is a common cause of death worldwide. With the aging of the population, the global burden of stroke is expected to increase [6]. When ischemic stroke occurs, the BBB can be disrupted, resulting in a number of impairments that can worsen the disease's impact. Currently, maintaining the integrity of the BBB is regarded as an effective treatment strategy for stroke [7].

Blood-brain barrier Composition of BBB

The neurovascular unit (NVU) serves as the anatomical basis for the BBB (Fig. 1). It is a tightly functioning cellular system consisting of endothelial cells, pericytes and astrocytes. In addition, it receives support from other types of central nervous system structures [8]. The growth and maintenance of BBB are controlled by the interaction between non-cellular elements and endothelial cells [9].



© The Author(s) 2023. Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativeco mmons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

^{*}Correspondence:

Jin-Xia Hu

jinxia0819@163.com

¹ Institute of Stroke Research, Xuzhou Medical University, Jiangsu, China ² Department of Neurology, The Affiliated Hospital of Xuzhou Medical

University, Jiangsu, China

³ School of Chemical Engineering and Technology, China University of Mining and Technology, Xuzhou 221116, China



Fig. 1 BBB structure

Neurovascular unit

The neurovascular unit (NVU) [10] is composed of a complex cellular structure, with each component having an intimate relationship that forms a highly efficient system for regulating cerebral blood flow [11]. The BBB is part of the NVU, which exists as a complex that combines vessels and astrocytes with neurons [12]. The NVU typically comprises encephalic vessels, emphasizing the close physical and functional connectivity between brain tissue and vessels [13].

Endothelial cells and tight junction (TJ)

Endothelial cells belong to the squamous cell family and are vital components that participate in forming the lining of blood vessels. Aside from their function in constructing tight junctions, endothelial cells also express specific transport proteins that regulate the dynamic flow of substrates. More importantly, endothelial cells can manage the transportation of leukocyte cell adhesion molecules to limit inflammatory invasion [14]. The TJ complex consists of transmembrane adhesive proteins that mediate intracellular signal transduction and provide necessary physical support by interacting with corresponding material on the adjacent cytoplasmic membrane [15].

Pericyte

Pericytes are smooth muscle-like cells that are distributed along capillaries and minute vessels. They possess contractility and are coated within the basement membrane, which plays an important role in maintaining nervous system function. During the period of angiogenesis and maturation of the BBB, pericytes regulate capillary vessel diameter, brain flow [16], and prevent immune cells from penetrating into the central nervous system [9, 17].

Astrocyte

In the central nervous system, astrocytes have various functions [18]. Their different polarization types possess diverse biochemistry and feature characteristics [19]. Astrocytes can offer neurons energy substrate [20], regulate local blood flow [21], help with drainage of interstitial fluid [22], hold synapse growth and plasticity [23], express benefits on function and behavior about neural circuits [24], and keep the balance state of extracellular fluid,ions, and neurotransmitters [25].

Microglia cell

Microglia cells are a type of cerebral congenital immune cell [26]. They can adjust tissue development, maintain neural environment stability, promote nerve-repairing procedures, and respond promptly to stressors [27]. Recently, much research indicates that activation of microglia cells is a critical factor in a variety of disease conditions, including ischemic stroke and Alzheimer's disease [28, 29].

Extracellular matrix

The extracellular matrix (ECM) is a layer of glycocalyx made up of proteoglycans and glycosaminoglycans that covers the BBB [30]. It functions as the first line of defense against substances attempting to cross the BBB [31]. The ECM proteins and receptors play a crucial role in various molecular signal transductions. They regulate cellular survival, development, metastasis, and differentiation, and enable the brain to adapt to

environmental changes [32, 33]. The ECM exists in all tissues and undergoes constant remodeling in response to control signals [33, 34]. Its dynamic structure allows it to regulate cellular signal transduction through integrins and other surface receptors. The ECM's relative elasticity and ability to sense mechanical signals help regulate organogenesis and adjust cellular metabolism [35]. Matrix metalloproteinases (MMPs) are members of the zinc-dependent endopeptidase family that play an important role in regulating the duration of ECM degradation and remodeling [36]. The expression level and functional changes of MMPs could trigger abnormal ECM degeneration, and are considered an initial signal of disease development [37, 38]. In some pathological conditions, ECM remodeling can become out of control, leading to conditions such as cancer. Excessive accumulation of ECM components is strongly associated with fibrosis [39]. When the BBB is destroyed, plasma protein leakage increases, causing protein retention and leading to adverse effects on the nervous system [40]. The precision of ECM manipulation stems from its composition of glycoproteins, which allows for the use of proteolytic cleavage to modulate the pathophysiological function of the BBB [41]. Currently, numerous methods and drugs are under investigation to modify, modulate, or mimic ECM components in order to effectively treat associated diseases. These efforts demonstrate a commitment to advancing medical research and developing innovative solutions for improved patient outcomes.

Biological function of the BBB

The BBB is a physical and biochemical barrier that precisely regulates environmental homeostasis in brain tissue. It provides an optimal microenvironment for neurons to perform their functions [42]. Under normal physiological conditions, the BBB ensures a constant supply of nutrients (such as oxygen and glucose) to cerebral cells and directs inflammatory cells to respond to local environmental changes [43-45]. The BBB regulates the microenvironment through its ion channels and transport proteins, maintaining the ion homeostasis and nutrients required by the brain. It also regulates the level of neurotransmitters in the brain and restricts the infiltration of plasma proteins and neurotoxins into the brain [46]. However, if the integrity of the BBB is compromised, it can result in ion imbalance, changes in signal molecule homeostasis, and attack by immune cells and molecules on the central nervous system, leading to neuronal dysfunction, degeneration, and various neurological diseases [47].

The TJ and junctional proteins of BBB

The TJ [48] primarily consists of occluding proteins and claudins. Among these, claudin-5 is recognized as the dominant TJ protein, making a significant contribution [49]. The zonula occludens (ZO) protein family, a group of membrane-associated guanylate kinase tight junctions, plays a central role in scaffold protein binding to the cytoskeleton [50]. Tight junctions significantly reduce the osmosis of polar solutes from plasma to extracellular fluid through the paracellular pathway [51]. Adhesion junctions [48] involve cadherin, platelet endothelial cell adhesion molecules, and JAM molecules of the adhesion family. Members of this family play an essential role in leukocyte adhesion and trans-BBB migration to the brain parenchyma [52]. However, the specific role of various binding complexes and related proteins in the development and physiological and pathological mechanisms of the BBB requires further clarification. Brain endothelial cells also have gap junctions formed by the connexin family [53], which allow intercellular communication and maintain the integrity of tight connections [48, 54].

The permeability and pathways through the BBB

The permeability of the BBB is determined by its structure, which is related to the interaction between molecular characteristics and transport proteins. The molecular characteristics that contribute to permeability include molecular weight, hydrogen bonding, polar surface area, electric charge, lipophilicity, and other factors [55–57].

There are two ways for materials to traverse the BBB: the transcellular pathway and the paracellular pathway, or, to put it more simply, across the cell itself or via paracellular space at the cell junctions (Fig. 2) [58]. The paracellular pathway is a passive transport mechanism regulated by osmotic pressure and concentration gradient. This process is highly restricted by the TJ and is limited to the diffusion of ions and small hydrophilic molecules [59, 60]. The transcellular pathway is mediated by various mechanisms and is divided into active and passive transport pathways depending on whether energy is consumed during transportation. Passive transport occurs through transcellular diffusion, while the active transcellular transport pathway includes receptor-mediated endocytosis, active efflux transport, and adsorption-mediated endocytosis [61]. Certain gases, like O2 and CO2, can passively diffuse across cells, as can small lipophilic molecules with Log P<5 and molecular weight < 500 Da [59, 62]. However, polar macromolecules such as peptides and proteins require receptor-mediated endocytosis for transport to their target locations and belong to the active transcellular transport pathway [56, 63, 64]. The BBB expresses various transport



Fig. 2 Transportation of substance on the BBB

proteins, including glucose, choline, and iron transporters, as well as insulin binding proteins and active efflux pumps composed of ATP-binding box transporters [65, 66]. ATP-binding box transporters, like p-glycoprotein, use ATP energy to prevent drugs, exogenous substances, neurotoxic substances, and nucleosides from actively flowing out of endothelial cells into the blood [54]. The transcellular transport pathway is more likely to cross the BBB than the paracellular pathway and is the focus of research for many drug delivery strategies across the BBB [60]. Adsorptive transcytosis relies on the nonspecific transport of positively charged substrates, such as cationic bovine serum albumin, interacting with the negatively charged surfaces of brain endothelial cells [67]. The integrity of the BBB is closely associated with the endothelial cell under physiological conditions; but it can be influenced by immune cells, such as microglia and macrophages, during pathological events [68]. Because pathological factors and physical and chemical stimuli can alter the BBB's permeability, numerous imaging technologies have been developed and applied to measure and evaluate changes in the BBB [63].

BBB injuries after ischemic stroke

The damage to the BBB is a crucial pathological process of ischemic stroke, which begins during the ischemic phase, worsens during the reperfusion stage, and ultimately results in vasogenic edema and hemorrhagic transformation (Fig. 3) [69–71]. Early-stage pathophysiological events associated with BBB breakdown after an ischemic stroke include the stimulation of sodium transporters (such as Na-K-Cl cotransporters, Na-H exchangers, etc.), which results in edema, and oxidative stress involving reactive oxygen species [72]. The degradation of integrin or TJ proteins also leads to an increase in paracellular leakage of the BBB [43]. Consistent damage to the BBB can cause neuroinflammation and the infiltration and accumulation of immune cells in the brain parenchyma [43].

Ischemic edema is a significant manifestation of ischemic stroke. The reasons for edema are changes in BBB permeability induced by ischemia that result in a series of cytotoxic, ionic, and vasogenic edema events [73–75]. The cell plasma membrane exhibits increased selective permeability, Na+/K+-ATPase, and Ca2+-ATPase activity, leading to the accumulation of sodium-dominated ions and water in cells as a precursor of ionic edema [73, 76, 77]. Many transport proteins close to the BBB may interfere with the stability of the adjustment mechanism on ion channel-associated transporters (Na-H exchangers, Na-Ca exchangers, Na-K-Cl cotransporters), which are non-selective cation channels controlled by sulfonylurea receptor-1 [78]. Vasogenic edema develops from ionic edema. The gaps in the capillaries'



Fig. 3 General pathophysiological phases and its main pathophysiological processes of ischemic stroke

basement membrane widen further due to the breakdown of tight connections between endothelial cells, allowing protein-rich fluid to seep from the extracellular fluid of the brain cells [73, 75].

After an ischemic stroke occurs, surrounding immune cells (such as monocytes, neutrophils, T cells, and others) and microglia work together to mediate the death of neurons and the breakdown of the BBB (Fig. 4) [79]. The attack increases the BBB's permeability, allowing immune cells and plasma proteins to enter the brain parenchyma, worsening the inflammatory process [72]. Microglia and

astrocytes activation following the ischemia leads to the production of cytokines in ischemic brain tissue, and these cytokines and matrix metal proteinases (MMPs) are critical mediators in BBB injury during ischemic stroke. They cause an increase in adhesion molecules and inflammatory blood cells, primarily neutrophils, which infiltrate through the damaged BBB [80]. Interleukin-1 β (IL-1 β) induces pericellular secretion of matrix metal-loproteinase-9 (MMP-9) through the NF- κ B signaling pathway, which damages the BBB's integrity [81, 82]. MMP-9 is involved in BBB injury pathogenesis through



Fig. 4 Neuroinflammatory mechanism and related factors of BBB ischemic stroke injury

the potential NOTCH3/NF-κB signaling pathway. The mitogen-activated protein kinase (MAPK) signaling pathway also plays a crucial role in ischemic strokes [83–85]. P38 MAPK regulates the synthesis of occludin proteins in the BBB structure [86]. Chemokines mediate secondary brain injury by activating MAPK-related signaling pathways, making targeting and restraining the p38 MAPK pathway vital to protect the BBB's structural integrity [87]. The Wnt/ β -catenin signaling pathway plays a critical role in the formation, maintenance, and development of the BBB [4, 88-92]. Activation of this pathway leads to an increase in the expression of several Wnt ligands such as Wnt-1, Wnt-3a, and Wnt-5 A, as well as β -catenin protein [93]. The Wnt/ β -catenin signaling pathway is involved in the regulation of central nervous system angiogenesis and the expression of BBB-specific transporter molecules, promoting the formation of capillary TJ proteins [94-96]. Moreover, Wnt5a can regulate endothelial cell survival, proliferation, and gene expression [97, 98]. The Wnt7a/7b ligand and Wnt/ β -catenin signaling pathways drive angiogenesis in the brain and BBB generation [99]. However, during ischemic stroke attack, the expression of Wnt-3a and β -catenin is downregulated, leading to BBB injury [93, 100]. Recent studies have demonstrated that microRNAs (miRNAs) play a crucial role in regulating changes in gene expression in brain microvascular endothelial cells associated with inflammation [101].

Major injury-related cytokines

interleukin-1 β (IL-1 β), tumor necrosis factor α (TNF- α), IL-6, IL-10, interferon β (IFN- β) and transforming growth factor β (TGF- β)), chemokines (e.g., Monocyte chemoattracted-protein 1 (MCP-1/CCL2), MIP-1 α (CCL3) and SDF-1 (CXCL12)), MMPs, and vascular endothelial growth factor (VEGF).

Apart from the immune neuroinflammatory response, oxidative stress also plays a role in cerebral ischemiareperfusion injury. These two factors interact to mediate neuron and BBB damage during ischemic stroke and subsequent hemorrhagic transformation (HT) [102]. Under physiological conditions, the oxidative stress process involves a series of peroxides and superoxides, including reactive oxygen species (ROS), reactive nitrogen species (RNS), and other reactive intermediates. These act as important regulators in the transmission of redox reaction signals. However, during ischemic stroke or other ischemia-hypoxia injuries, the physiological balance mechanism between ROS/RNS production and elimination is disrupted, leading to oxidative/nitrosative stress and persistent oxidative damage [103]. This disruption also affects electron transport chains and mitochondrial respiration, disrupting mitochondrial dynamics and ATP

synthesis [104], which can injure neurons, activate the apoptosis pathway, and further lead to oxidative damage of BBB endothelial cells [105]. Reactive oxygen species (ROS) are key mediators of BBB dysfunction during oxidative stress, regulating TJ proteins and the cytoskeleton of brain endothelial cells [72]. They participate in oxidative damage, regulate TJ modification, and activate inflammatory factors, and thus play a crucial role in the various mechanisms of BBB damage, including the kinin system, excitatory toxicity from toxic glutamate efflux, neutrophil recruitment, mitochondrial changes, and macrophage/microglial activation [106, 107]. Glutathione (GSH) is another important participant in the REDOX process, and its oxidation to glutathione disulfide (GSSH) is a critical step. Interference with this REDOX metabolism may impair barrier homeostasis and produce oxidative stress [108, 109].

As mentioned earlier, when an ischemic stroke occurs in the brain, a cascade of inflammatory mediators (including cytokines, chemokines, and growth factors) is released into the damaged tissues [110, 111]. These inflammatory mediators can cause the release of MMPs, which can break down the TJ proteins and compromise the integrity of the BBB [112, 113]. TJ proteins, such as the claudin family, occludin, Zona occludens 1 (ZO-1), and tricellulin, have been identified as being related to the neuro-barrier [114]. The mechanism of BBB injury after an ischemic stroke lies in the early up-regulation of endocytosis in endothelial cells and the later remodeling of tightly connected complexes [115]. Increasing evidence suggests that the primary mechanism behind BBB leakage after a stroke is the breakdown of the TJ complex. Integrin, which influences cell adhesion, migration, and survival, has been found to have a significant impact on these processes [116]. Laminin is an ECM protein that is widely expressed in the CNS and can interact with both integrin and non-integrin receptors [117]. Numerous studies have explored the relationship between integrin and BBB permeability. After an ischemic stroke, the upregulation of integrin $\alpha v\beta 3$ can promote angiogenesis and functional recovery [118]. Additionally, the induction of integrin $\alpha 5\beta 1$ and its downstream signaling pathway play a crucial role in the pathology of ischemic stroke and cerebral hypoxia [119]. However, there is still some controversy over the expression and specific roles of laminin and various integrins during ischemic stroke [117].

Furthermore, reperfusion-induced hemorrhagic transformation is a common complication after ischemic stroke. During the initial period, leukocyte-derived MMP-9 and brain-derived MMP-2 are involved, and later, MMP-3 and MMP-9, angiogenesis, and vasogenic edema may occur, which can destroy NVUs and exacerbate the destruction of the BBB [120, 121].

Treatment approaches

Currently, recombinant tissue-type plasminogen activator (r-tPA) thrombolytic therapy remains the most effective treatment for ischemic stroke. However, due to the narrow time window of 4–6 h after stroke onset, only a small number of patients can receive efficient thrombolytic therapy. This limited time frame poses a challenge for stroke treatment [122], and the risk of hemorrhagic transformation after application further restricts its clinical application [123].

Improve cerebral edema

Early surgical decompression is a significant factor in improving the prognosis of cerebral edema caused by an ischemic stroke. Although therapeutic hypothermia is a potential treatment option, it remains unproven, and conservative drugs have limited efficacy in anti-edema treatment [124]. Targeting caspase-1 has been shown to reduce cerebral edema and the incidence of hemorrhagic transformation (HT) during acute stroke [125]. Caspase-1 is a family of cysteine proteases that mediate pyroptosis [126]. In acute stroke, caspase-1 is upregulated and has been shown to mediate BBB disruption [127].

Regulates immune and inflammatory responses

Regulating immune and inflammatory responses is crucial in treating ischemic stroke. Immune cells can help eliminate necrotic tissue and promote neuron recovery, but they also release inflammatory factors that aggravate breakdown of the BBB, especially during later reperfusion [128]. Targeting immune cells in BBB disruption is a promising strategy to improve stroke prognosis and existing treatments [129]. Currently, immunoregulatory therapies are being developed to reduce pro-inflammatory cytokines, MMPs, and infiltrating leukocytes to maintain BBB homeostasis, although there are no proven clinical applications for immunoregulation yet. Ischemic stroke triggers a serious neuroinflammatory response [128, 129], which can harm neurons by releasing cytokines, chemokines, and oxidative stress-related factors. Therefore, taking corresponding measures to suppress the immune response and the occurrence and development of inflammatory processes during cerebral ischemia may be a promising target for developing new therapeutic strategies [130]. Several methods are being studied to inhibit MMPs, which mediate TJ destruction and protect the BBB from ischemic injury. Physical methods like hyperbaric oxygen, hypothermia, and drugs like isoflurane and hydrogen sulfide, or non-invasive vagus nerve stimulation, can all be used to achieve this goal [131, 132]. Activated astrocytes play an important role in a series of inflammatory reactions after an ischemic stroke. Li [133] reviewed the possibility of targeting multiple reactive astrocytes to protect the BBB and maintain brain homeostasis. Qu [134] found that gallic acid (GA) can alter microglia polarization to reduce BBB injury induced by cerebral ischemia/reperfusion with beneficial consequences.

Eliminate oxidative stress

Mitochondrial dynamics, which include ROS generation, autophagy, and cell apoptosis, are closely linked to the pathophysiology of ischemic stroke. These dynamics also affect the body's energy metabolism. Some researchers believe that inhibiting excessive mitochondrial division and restoring the balance of mitochondrial dynamics could be a novel approach to treating ischemic stroke [135]. Certain natural polyphenols act as powerful antioxidants that inhibit ROS production, scavenge free radicals, and improve BBB function [136]. Additionally, intravenous administration of the endogenous peptide apelin-13 can significantly reduce BBB permeability and vasogenic edema by targeting oxidative stress during ischemia-reperfusion.

Reconstruct the BBB structural components and regulate the signaling pathway

Reconstructing the BBB is considered a promising treatment option for ischemic stroke. Kadir RRA et al. [137] established an in vitro BBB model through cell co-culture and demonstrated that overgrown endothelial cells (OECs) can effectively migrate to the injured site and restore BBB integrity. OEC-based cell therapy can also reduce oxidative stress and apoptosis of cerebral microvascular endothelial cells after ischemic stroke injury. M. Alwjwaj [138] later found that OEC acts as a therapeutic agent to prevent ischemia by specifically inhibiting NOX2, a major source of vascular oxidative stress, and thereby reducing oxidative stress. Zeng et al. [139] conducted in vivo and in vitro experiments and found that the DNA methyltransferase inhibitor Zebularine can reduce the production of pro-inflammatory factors and improve brain edema and nerve function by increasing the expression of ZO-1 and vascular endothelial (VE)cadherin [131, 132]. Nilles KL reviewed [140] efforts to optimize the success rate of stroke drug conversion across the BBB by targeting BBB transporters. Song et al. found that selective loss of Nhe1 (a Ph-sensitive Na + /H + exchanger 1) in astrocytes can increase the expression of Wnt7a/7b protein and preserve Wnt/βcatenin signal in endothelial cells, improve angiogenic repair, cerebral blood perfusion, and maintain the integrity of the BBB after ischemic stroke [141]. Recent studies have also been conducted to synthesize specific Wnt activators, such as Wnt7a, through genetic engineering

to protect the BBB [142]. Studies suggest that Fluoxetine can increase the expression of Claudin-5 and its effect on Wnt signaling may help restore BBB function while promoting neurogenesis [143, 144]. Lithium can lower the expression of MMP-9, increase the activity of Wnt/ β -catenin signaling, and preserve the integrity of the BBB by raising the TJ protein levels (Claudin-5 and ZO-1) [145]. Laksitorini et al. reported that activation of Wnt/β-catenin by LiCl (GSK3 inhibitor) or Wnt3a can improve brain endothelial barrier function in the BBB cell culture model in vitro [144]. Liu et al. demonstrated that the glucagon-like peptide-1 receptor (GLP-1R) agonist exendin-4 (EX-4) inhibits ROS production and MMP-9 activation by activating the Wnt/β-catenin signaling pathway in a rat model of temporary middle cerebral artery occlusion. Therefore, GLP-1R agonists can be potential protective agents to reduce the risk of HT after r-tPA treatment [146]. Golgi matrix protein 130 (GM130) [113]can up-regulate the inhibition of the autophagylysosome pathway, maintain appropriate autophagy to prevent the damage of tight connections, maintain BBB function, and reduce brain parenchymal injury. Exosomes isolated from the venous serum of healthy individuals can also play a neuroprotective role against experimental stroke by inhibiting endothelial cell apoptosis and BBB breakdown mediated by autophagy [147, 148]. This approach shows promise as a potential treatment for ischemic stroke through the BBB in the future.

The application of novel nanomaterials

The BBB is a significant obstacle to drug delivery to the brain parenchyma [149]. However, this barrier can be overcome by opening or crossing the endothelial barrier, and specific sites can be targeted to protect corresponding organs [150]. Therefore, there is growing interest in studying the delivery of neuroprotective agents to enable drugs to cross the BBB and reach their target sites in the brain parenchyma. Nanoparticles such as liposomes have been used for drug delivery, as they can minimize chemical degradation, improve drug permeability, and achieve the necessary blood drug concentration at the site of action [151–153]. For instance, cerium (Ce)-doped Linde Type A (LTA) zeolite-based nanomaterials (Ce/ Zeo-NMs) have been shown to use their unique adsorption capacity and simulated catalytic activity to remove reactive oxygen species, enhance the integrity of the BBB, inhibit the activation of inflammatory cells, and reduce neurovascular dysfunction [154]. Betulinic acid, one of the most potent stroke antioxidants, can be delivered by naturally compound-derived nanoparticles (NPs) to improve stroke recovery [155]. Moreover, nanomaterials coated with drugs, such as uPA-loaded black phosphorus nanosheets (BPNs), can effectively deliver uPA across the BBB to dissolve thrombi and remove ROS [156]. Braintargeting bionic nanomaterials (RR@SABNPs) can significantly prolong the half-life of salvianolic acid B (SAB), deliver SAB to the ischemic brain, and demonstrate a good therapeutic effect in model mice [157]. The nanogel system is a promising drug delivery platform that could be used therapeutically as a more stable and superior option for crossing the BBB [140, 158]. Invasive techniques such as ultrasound drug delivery, craniotomy drug delivery, and other methods are related to brain drug delivery strategies targeting intracranial diseases [155, 159].

Chinese medicine treatment

Currently, it has been discovered that some traditional Chinese medicines or prescriptions can protect against ischemic stroke through various mechanisms. The Qishen Yiqi formula [160] protects against ischemic stroke through a synergistic lysosomal/inflammatory mechanism. Certain Chinese herbs [161] can enhance their thrombolytic ability and reduce the risk of hemorrhagic transformation as tPA adjuncts. Astragaloside IV (AST IV) and total saponins of notoginseng (PNS) are the main effective components of Astragaloside IV and notoginseng for treating ischemic stroke, respectively. When combined with borneol, these components can promote their delivery by down-regulating the expression of effector transporters and up-regulating the expression of uptake transporters, thereby enhancing the protective effect of ischemia-reperfusion and maintaining BBB integrity [162]. Moreover, other studies have shown that acupuncture can activate the inherent antioxidant enzyme system, inhibit the overgeneration of reactive oxygen species, reduce the potential of oxidative stress caused by cerebral ischemia, and play a neuroprotective role [163, 164].

Other methods

Clinical trials have suggested that stem cell therapy holds promise in improving the sequelae of patients with acute ischemic stroke, although further verification is still needed. Neural stem cell transplantation, which can replace damaged neural cells and maintain the BBB through the bystander effect, is a potential therapy for neurovascular diseases [165, 166]. In recent years, microRNA has also received significant research attention due to its negative impact on BBB injury after ischemic stroke. Intervening with microRNA can control BBB permeability through various mechanisms [131, 167]. Further research indicates that microRNA could serve as a promising BBB modulator for ischemic brain injury. Another study found that the activation of vascular endothelial growth factor (VEGF) can strengthen the



Fig. 5 Graphic abstract of this article

vasculature in stroke patients and prevent the progression of secondary brain damage. Furthermore, administering VEGF after a stroke can repair the BBB and reduce secondary brain edema damage [168].

What's more, the application of single-cell sequencing in the analysis and collection of genetic information related to clinical diseases is increasingly prevalent. Among them, RNA sequencing (RNA-Seq) is widely used and has gradually become an indispensable tool for analyzing differential gene expression. Based on this, a large number of RNA sequences have also been introduced into BBB research in recent years. The earliest genom-FIGHic analysis of BBB components using RNA-Seq identified that genes related to tight junctions, signaling molecules, and some other molecular genes are highly enriched at the BBB, indicating the involvement of Wnt and RXRalpha in the signaling cascade regulation of this barrier [169]. David and his team summarized relevant scientific experience to establish guidelines that promote RNA-seq research on the BBB [170]. He and his team captured corresponding vascular cells using transgenic reporter mice, thus constructing a database of mouse brain and lung vascular and vascular-related cell types. This database will provide a solid foundation for vascular development and disease research [171]. There are also some other relevant studies that compare the transcriptional gene expression differences between human and mouse brain micro-vessels, revealing their potential impact on drug delivery and diseases. After all, increasing the understanding of gene expression differences between the mouse and human brain vascular systems is crucial to assessing the potential limitations of mouse models for

studying brain vasculature and the BBB development and diseases [172]. Furthermore, through single-cell sequencing and gene expression analysis of differential results, it is possible to guide experimental research in the search for disease target factors. Recently, a study using RNA-Seq has provided a detailed comparative characterization of the molecular profiles of endothelial cells in normal human brain tissue and glioblastoma, aiming to provide valuable information for drug delivery across the BBB and intra-tumoral distribution [173]. Other studies have used RNA-seq to explore potential targets and signaling pathways related to BBB dysfunction induced by A1 astrocytes. It is inferred that blocking the transformation of C3d+/GFAP+A1 astrocytes may alleviate BBB disruption in mice after ischemic stroke [174] (Fig. 5).

Conclusion and expectation

By providing a summary of our review, we aim to offer new insights for enhancing the prognosis of future research on the treatment of ischemic stroke by safeguarding the integrity of the BBB. We are confident that, with the collective efforts of an increasing number of researchers, more effective and innovative approaches will be developed and applied in clinical settings to alleviate the social and economic burden that this disease places on families.

Abbreviations

BBB	Blood brain barrier
ATP	Adenosine triphosphate
IL-1β	Interleukin-1β
MAPK	Mitogen-activated protein kinase
HT	Hemorrhagic transformation

ROS	Reactive oxygen species
RNS	Reactive nitrogen species
GSH	Glutathione
GSSH	Glutathione disulfide
REDOX	Reduction-oxidation
r-tPA	Recombinant tissue-type plasminogen activator
GA	Gallic acid
OECs	Overgrown endothelial cells
NOX2	NADPH oxidase 2
GLP-1R	Glucagon-like peptide-1 receptor
GM130	Golgi matrix protein 130
Ce/Zeo-NMs cerium	(Ce)-doped Linde Type A (LTA) zeolite-based nanomaterials
NPs	Nanoparticles
uPA	Urokinase-type plasminogen activator
BPNs	Black phosphorus nanosheets
SAB	Salvianolic acid B
AST IV	Astragaloside IV
PNS	Saponins of notoginseng
VEGF	Vascular endothelial growth factor

Acknowledgements

Not applicable.

Author contributions

G.H.M and C.H. contributed to the manuscript preparation. G.Y.C and J.X.H. contributed to the revision of the manuscript. All authors read and approved the final manuscript.

Funding

This research was supported by the National Natural Science Foundation of China (82001276, 82171305), the Fundamental Research Funds for the Central Universities (Grant No. 2018BSCXA12) and Postgraduate Research & Practice Innovation Program of Jiangsu Province (Grant No. KYCX18_1931).

Availability of data and materials

Not applicable.

Declarations

Ethics approval and consent to participate Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare no competing interests.

Received: 11 June 2023 Accepted: 4 September 2023 Published online: 01 November 2023

References

- Ehrlich P. Farben-therapeutische Versuche bei Trypanosomerkankung. Berliner Klin Wochenschr. 1904;14:362-5
- Goldmann EE. Vitalfärbung am Zentralnervensystem: Beitrag zur Physio-Pathologie des Plexus chorioideus und der Hirnhäute. Königl. Akademie der Wissenschaften; 1913.
- Stern L, Gautier R. Recherches sur Le liquide céphalo-rachidien: I.–Les rapports entre Le liquide céphalo-rachidien et la circulation sanguine. Archives internationales de physiologie. 1921;17(2):138–92.
- Liebner S, et al. Functional morphology of the blood-brain barrier in health and disease. Acta Neuropathol. 2018;135(3):311–36.
- Lozano Villanueva JL, et al. Association between heart failure and clinical prognosis in patients with acute ischemic stroke: a retrospective cohort study. J Clin Neurol. 2021;17(2):200–5.

- Benjamin EJ, et al. Heart disease and stroke statistics-2018 update: a report from the American heart association. Circulation. 2018;137(12):e67–492.
- 7. Jiang X, et al. Blood-brain barrier dysfunction and recovery after ischemic stroke. Prog Neurobiol. 2018;163–4:144–71.
- Marta S, Maria RA, Amparo A-P. Blood–brain barrier dynamics to maintain brain homeostasis. Trends Neurosci. 2021;44(5):393–405.
- Obermeier B, Daneman R, Ransohoff RM. Development, maintenance and disruption of the blood-brain barrier. Nat Med. 2013;19(12):1584–96.
- Barichello T et al. An overview of the blood-brain barrier. Blood-Brain Barrier, 2019: p. 1–8.
- 11. Abbott NJ, Friedman A. Overview and introduction: the blood-brain barrier in health and disease. Epilepsia. 2012;53(0 6):1–6.
- 12. Muoio V, Persson PB, Sendeski MM. The neurovascular unit concept review. Acta Physiol (Oxf). 2014;210(4):790–8.
- 13. Keller A. Breaking and building the wall: the biology of the blood-brain barrier in health and disease. Swiss Med Wkly. 2013;143:w13892.
- Langen UH, Ayloo S, Gu C. Development and cell biology of the bloodbrain barrier. Annu Rev Cell Dev Biol. 2019;35:591–613.
- Bazzoni G, Dejana E. Endothelial cell-to-cell junctions: molecular organization and role in vascular homeostasis. Physiol Rev. 2004;84(3):869–901.
- 16. Attwell D, et al. What is a pericyte? J Cereb Blood Flow Metab. 2016;36(2):451–5.
- Zheng Z, Chopp M, Chen J. Multifaceted roles of pericytes in central nervous system homeostasis and disease. J Cereb Blood Flow Metabolism. 2020;40(7):1381–401.
- Sofroniew MV. Astrocyte reactivity: subtypes, states, and functions in CNS innate immunity. Trends Immunol. 2020;41(9):758–70.
- Gundersen GA, et al. Evidence that pericytes regulate aquaporin-4 polarization in mouse cortical astrocytes. Brain Struct Funct. 2014;219(6):2181–6.
- Magistretti PJ, Allaman I. Lactate in the brain: from metabolic endproduct to signalling molecule. Nat Rev Neurosci. 2018;19(4):235–49.
- 21. MacVicar BA, Newman EA. Astrocyte regulation of blood flow in the brain. Cold Spring Harb Perspect Biol. 2015;7(5):a020388.
- Plog BA, Nedergaard M. The glymphatic system in central nervous system health and disease: past, present, and future. Annu Rev Pathol. 2018;13:379–94.
- Allen NJ, Eroglu C. Cell biology of astrocyte-synapse interactions. Neuron. 2017;96(3):697–708.
- 24. Khakh BS. Astrocyte-neuron interactions in the striatum: insights on identity, form, and function. Trends Neurosci. 2019;42(9):617–30.
- Verkhratsky A, Nedergaard M. Physiology Astroglia. Physiol Rev. 2018;98(1):239–389.
- Sierra A, et al. The "Big-Bang" for modern glial biology: translation and comments on Pío del Río-Hortega series of papers on microglia. Glia. 2016;64(11):1801–40.
- Subramaniam SR, Federoff HJ. Targeting microglial activation states as a therapeutic avenue in Parkinson's disease. Front Aging Neurosci. 2017;9:176.
- Ronaldson PT, Davis TP. Regulation of blood-brain barrier integrity by microglia in health and disease: a therapeutic opportunity. J Cereb Blood Flow Metab. 2020;40(1suppl):S6–s24.
- Gupta N, et al. Recent progress in therapeutic strategies for microgliamediated neuroinflammation in neuropathologies. Expert Opin Ther Targets. 2018;22(9):765–81.
- Reed MJ, Damodarasamy M, Banks WA. The extracellular matrix of the blood-brain barrier: structural and functional roles in health, aging, and Alzheimer's disease. Tissue Barriers. 2019;7(4):1651157.
- 31. Logsdon AF, et al. The neurovascular extracellular matrix in health and disease. Exp Biol Med (Maywood). 2021;246(7):835–44.
- 32. Theocharis AD, et al. Extracellular matrix structure. Adv Drug Deliv Rev. 2016;97:4–27.
- Bonnans C, Chou J, Werb Z. Remodelling the extracellular matrix in development and disease. Nat Rev Mol Cell Biol. 2014;15(12):786–801.
- 34. Hynes RO. The extracellular matrix: not just pretty fibrils. Science. 2009;326(5957):1216–9.
- 35. Manou D, et al. The complex interplay between extracellular matrix and cells in tissues. Methods Mol Biol. 2019;1952:1–20.

- 36. Singh D, et al. Multifaceted role of matrix metalloproteinases (MMPs). Front Mol Biosci. 2015;2:19.
- 37. Cabral-Pacheco GA, et al. The roles of matrix metalloproteinases and their inhibitors in human diseases. Int J Mol Sci. 2020;21(24):9739.
- 38. Rezvan A, et al. CD147 and MMPs as key factors in physiological and pathological processes. Biomed Pharmacother. 2023;157:113983.
- Kyriakopoulou K, et al. Trends in extracellular matrix biology. Mol Biol Rep. 2022;50:853.
- Baeten KM, Akassoglou K. Extracellular matrix and matrix receptors in blood-brain barrier formation and stroke. Dev Neurobiol. 2011;71(11):1018–39.
- Thomsen MS, Routhe LJ, Moos T. The vascular basement membrane in the healthy and pathological brain. J Cereb Blood Flow Metab. 2017;37(10):3300–17.
- Praveen B, Alex B, Maiken N. The blood-brain barrier: an overview: structure, regulation, and clinical implications. Neurobiol Dis. 2004;16(1):1–13.
- Abdullahi W, Tripathi D, Ronaldson PT. Blood-brain barrier dysfunction in ischemic stroke: targeting tight junctions and transporters for vascular protection. Am J Physiol Cell Physiol. 2018;315(3):C343–C356.
- Erickson MA, Banks WA. Neuroimmune Axes of the blood-brain barriers and blood-brain interfaces: bases for physiological regulation, Disease States, and pharmacological interventions. Pharmacol Rev. 2018;70(2):278–314.
- Persidsky Y, et al. Blood-brain barrier: structural components and function under physiologic and pathologic conditions. J Neuroimmune Pharmacol. 2006;1(3):223–36.
- Kadry H, Noorani B, Cucullo L. A blood-brain barrier overview on structure, function, impairment, and biomarkers of integrity. Fluids Barriers CNS. 2020;17(1):69.
- 47. Daneman R, Prat A. The blood-brain barrier. Cold Spring Harb Perspect Biol. 2015;7(1):a020412.
- Zhao Z, et al. Establishment and dysfunction of the blood-brain barrier. Cell. 2015;163(5):1064–78.
- Berndt P, et al. Tight junction proteins at the blood–brain barrier: far more than claudin-5. Cell Mol Life Sci. 2019;76(10):1987–2002.
- Keep RF, et al. Brain endothelial cell junctions after cerebral hemorrhage: changes, mechanisms and therapeutic targets. J Cereb Blood Flow Metab. 2018;38(8):1255–75.
- Hartl N, Adams F, Merkel OM. From adsorption to covalent bonding: apolipoprotein E functionalization of polymeric nanoparticles for drug delivery across the blood–brain barrier. Adv Ther. 2021;4(1):2000092.
- Kummer D, Ebnet K. Junctional adhesion molecules (JAMs): the JAMintegrin connection. Cells. 2018;7(4):25.
- Stamatovic SM, et al. Junctional proteins of the blood-brain barrier: new insights into function and dysfunction. Tissue Barriers. 2016;4(1):e1154641.
- Sweeney MD, et al. Blood-brain barrier: from physiology to disease and back. Physiol Rev. 2019;99(1):21–78.
- Chowdhury EA, et al. Understanding the brain uptake and permeability of small molecules through the BBB: a technical overview. J Cereb Blood Flow Metabol. 2021;41(8):1797–820.
- 56. Kaya M, Ahishali B. Basic physiology of the blood-brain barrier in health and disease: a brief overview. Tissue Barriers. 2021;9(1):1840913.
- 57. Moody DM. The blood-brain barrier and blood-cerebral spinal fluid barrier. Semin Cardiothorac Vasc Anesth. 2006;10(2):128–31.
- Dong X. Current strategies for brain drug delivery. Theranostics. 2018;8(6):1481–93.
- 59. Whelan R, Hargaden GC, Knox AJS. Modulating the blood-brain barrier: a comprehensive review. Pharmaceutics. 2021;13(11):1980.
- Moura RP, et al. Blood-brain barrier receptors and transporters: an insight on their function and how to exploit them through nanotechnology. Expert Opin Drug Deliv. 2019;16(3):271–85.
- 61. Sarvari S, et al. Mechanisms in blood-brain barrier opening and metabolism-challenged cerebrovascular ischemia with emphasis on ischemic stroke. Metab Brain Dis. 2020;35(6):851–68.
- 62. Lipinski CA, et al. Experimental and computational approaches to estimate solubility and permeability in drug discovery and development settings. Adv Drug Deliv Rev. 2001;46(1–3):3–26.
- Fu BM. Transport across the blood-brain barrier. Adv Exp Med Biol. 2018;1097:235–59.

- 64. Terstappen GC, et al. Strategies for delivering therapeutics across the blood-brain barrier. Nat Rev Drug Discov. 2021;20(5):362–83.
- Xie J, et al. Nanomaterial-based blood-brain-barrier (BBB) crossing strategies. Biomaterials. 2019;224:119491.
- Li W, Sharma M, Kaur P. The DrrAB efflux system of Streptomyces peucetius is a multidrug transporter of broad substrate specificity. J Biol Chem. 2014;289(18):12633–46.
- 67. Georgieva JV, Hoekstra D, Zuhorn IS. Smuggling drugs into the brain: an overview of ligands targeting transcytosis for drug delivery across the blood-brain barrier. Pharmaceutics. 2014;6(4):557–83.
- Osipova ED, et al. Gliotransmitters and cytokines in the control of blood-brain barrier permeability. Rev Neurosci. 2018;29(5):567–91.
- Rousselet E, et al. Sustained (S)-roscovitine delivery promotes neuroprotection associated with functional recovery and decrease in brain edema in a randomized blind focal cerebral ischemia study. J Cereb Blood Flow Metabolism. 2018;38(6):1070–84.
- Turner RJ, Sharp FR. Implications of MMP9 for blood brain barrier disruption and hemorrhagic transformation following ischemic stroke. Front Cell Neurosci. 2016;10:56.
- Mracsko E, Veltkamp R. Neuroinflammation after intracerebral hemorrhage. Front Cell Neurosci. 2014;8:388.
- 72. Takata F, et al. Blood-brain barrier dysfunction amplifies the development of neuroinflammation: understanding of cellular events in brain microvascular endothelial cells for prevention and treatment of BBB dysfunction. Front Cell Neurosci. 2021;15:661838.
- 73. Kahle KT, et al. Molecular mechanisms of ischemic cerebral edema: role of electroneutral ion transport. Physiology. 2009;24(4):257–65.
- 74. Simard JM, et al. Brain oedema in focal ischaemia: molecular pathophysiology and theoretical implications. Lancet Neurol. 2007;6(3):258–68.
- 75. Turner RE, et al. High-altitude cerebral edema: its own entity or endstage acute mountain sickness? J Appl Physiol. 2021;131(1):313–25.
- Stokum JA, Gerzanich V, Simard JM. Molecular pathophysiology of cerebral edema. J Cereb Blood Flow Metabolism. 2016;36(3):513–38.
- 77. O'Donnell ME. Blood-brain barrier na transporters in ischemic stroke. Adv Pharmacol. 2014;71:113–46.
- 78. Shah K, Abbruscato T. The role of blood-brain barrier transporters in pathophysiology and pharmacotherapy of stroke. Curr Pharm Des. 2014;20(10):1510–22.
- 79. Ma Y, et al. The role of immune cells in post-stroke angiogenesis and neuronal remodeling: the known and the unknown. Front Immunol. 2021;12:784098.
- Yang C, et al. Neuroinflammatory mechanisms of blood-brain barrier damage in ischemic stroke. Am J Physiol Cell Physiol. 2019;316(2):C135–c153.
- Smyth LCD, et al. Unique and shared inflammatory profiles of human brain endothelia and pericytes. J Neuroinflammation. 2018;15(1):138.
- Yang F, et al. ATP induces disruption of tight junction proteins via IL-1 beta-dependent MMP-9 activation of human blood-brain barrier in vitro. Neural Plast. 2016;2016:8928530.
- Qin W, et al. Melatonin protects blood-brain barrier integrity and permeability by inhibiting matrix metalloproteinase-9 via the NOTCH3/ NF-κB pathway. Aging. 2019;11(23):11391–415.
- Song Y, et al. Activation of p38-mitogen-activated protein kinase contributes to ischemia reperfusion in rat brain. Genet Mol Res. 2016;15:1–3.
- Mohamed IN, et al. Role of inflammasome activation in the pathophysiology of vascular diseases of the neurovascular unit. Antioxid Redox Signal. 2015;22(13):1188–206.
- Ni Y, et al. TNFα alters occludin and cerebral endothelial permeability: role of p38MAPK. PLoS ONE. 2017;12(2):e0170346.
- Guo F, et al. Chemokine CCL2 contributes to BBB disruption via the p38 MAPK signaling pathway following acute intracerebral hemorrhage. Faseb j. 2020;34(1):1872–84.
- Chang J, et al. Gpr124 is essential for blood-brain barrier integrity in central nervous system disease. Nat Med. 2017;23(4):450–60.
- Lengfeld JE, et al. Endothelial Wnt/β-catenin signaling reduces immune cell infiltration in multiple sclerosis. Proc Natl Acad Sci. 2017;114(7):E1168–77.

- 90. Cottarelli A, et al. Fgfbp1 promotes blood-brain barrier development by regulating collagen IV deposition and maintaining Wnt/ β -catenin signaling. Development. 2020;147(16):dev185140.
- Corada M, et al. Fine-tuning of Sox17 and canonical wnt coordinates the permeability properties of the blood-brain barrier. Circul Res. 2019;124(4):511–25.
- Jia L, Piña-Crespo J, Li Y. Restoring Wnt/β-catenin signaling is a promising therapeutic strategy for Alzheimer's disease. Mol Brain. 2019;12(1):1–11.
- Chen X-Y, et al. Inhibition of the immunoproteasome LMP2 ameliorates ischemia/hypoxia-induced blood–brain barrier injury through the Wnt/β-catenin signalling pathway. Military Med Res. 2021;8(1):1–16.
- 94. Daneman R, et al. Wnt/ β -catenin signaling is required for CNS, but not non-CNS, angiogenesis. Proc Natl Acad Sci. 2009;106(2):641–6.
- Liu L, et al. Dysfunctional Wnt/β-catenin signaling contributes to blood-brain barrier breakdown in Alzheimer's disease. Neurochem Int. 2014;75:19–25.
- 96. Liebner S, et al. Wht/beta-catenin signaling controls development of the blood-brain barrier. J Cell Biol. 2008;183(3):409–17.
- Goodwin AM, Sullivan KM, D'Amore PA. Cultured endothelial cells display endogenous activation of the canonical wnt signaling pathway and express multiple ligands, receptors, and secreted modulators of wnt signaling. Dev Dyn. 2006;235(11):3110–20.
- Zerlin M, Julius MA, Kitajewski J. Wnt/Frizzled signaling in angiogenesis. Angiogenesis. 2008;11(1):63–9.
- 99. Engelhardt B, Liebner S. Novel insights into the development and maintenance of the blood-brain barrier. Cell Tissue Res. 2014;355(3):687–99.
- Qi C, et al. Hypoxia stimulates neural stem cell proliferation by increasing HIF-1α expression and activating Wnt/β-catenin signaling. Cell Mol Biol. 2017;63(7):12.
- Bernstein DL, et al. miR-98 reduces endothelial dysfunction by protecting blood-brain barrier (BBB) and improves neurological outcomes in mouse ischemia/reperfusion stroke model. J Cereb Blood Flow Metab. 2020;40(10):1953–65.
- 102. Chen H, et al. Therapeutic targets of oxidative/nitrosative stress and neuroinflammation in ischemic stroke: applications for natural product efficacy with omics and systemic biology. Pharmacol Res. 2020;158:104877.
- 103. Lehner C, et al. Oxidative stress and blood-brain barrier dysfunction under particular consideration of matrix metalloproteinases. Antioxid Redox Signal. 2011;15(5):1305–23.
- Gu M, Mei XL, Zhao YN. Sepsis and cerebral dysfunction: BBB damage, neuroinflammation, oxidative stress, apoptosis and autophagy as key mediators and the potential therapeutic approaches. Neurotox Res. 2021;39(2):489–503.
- Gyawali B, Ramakrishna K, Dhamoon AS. Sepsis: the evolution in definition, pathophysiology, and management. SAGE Open Med. 2019;7:2050312119835043.
- Pun PB, Lu J, Moochhala S. Involvement of ROS in BBB dysfunction. Free Radic Res. 2009;43(4):348–64.
- Yang J, et al. Modulation of vascular integrity and neuroinflammation by peroxiredoxin 4 following cerebral ischemia-reperfusion injury. Microvasc Res. 2021;135:104144.
- 108. Huang SF, et al. Astrocyte glutathione maintains endothelial barrier stability. Redox Biol. 2020;34:101576.
- Namba K, et al. Temporal profiles of the levels of endogenous antioxidants after four-vessel occlusion in rats. J Neurosurg Anesthesiol. 2001;13(2):131–7.
- Zhao B, et al. Research progress of mechanisms for tight junction damage on blood-brain barrier inflammation. Arch Physiol Biochem. 2020;128(6):1579–90.
- 111. Abdul-Muneer PM, et al. Role of matrix metalloproteinases in the pathogenesis of traumatic brain injury. Mol Neurobiol. 2016;53(9):6106–23.
- 112. Chen CY, et al. miR-195 reduces age-related blood-brain barrier leakage caused by thrombospondin-1-mediated selective autophagy. Aging Cell. 2020;19(11):e13236.
- 113. Deng S, et al. GM130 protects against blood-brain barrier disruption and brain injury after intracerebral hemorrhage by regulating autophagy formation. Exp Gerontol. 2022;163:111772.
- 114. Reinhold AK, Rittner HL. Barrier function in the peripheral and central nervous system-a review. Pflugers Arch. 2017;469(1):123–34.

- 116. Hamidi H, Ivaska J. Every step of the way: integrins in cancer progression and metastasis. Nat Rev Cancer. 2018;18(9):533–48.
- 117. Nirwane A, Yao Y. Laminins and their receptors in the CNS. Biol Rev Camb Philos Soc. 2018. https://doi.org/10.1111/brv.12454
- Bi JJ, Yi L. Effects of integrins and integrin αvβ3 inhibitor on angiogenesis in cerebral ischemic stroke. J Huazhong Univ Sci Technolog Med Sci. 2014;34(3):299–305.
- 119. Edwards DN, et al. Integrin α 5 β 1 inhibition by ATN-161 reduces neuroinflammation and is neuroprotective in ischemic stroke. J Cereb Blood Flow Metab. 2020;40(8):1695–708.
- Jickling GC, et al. Hemorrhagic transformation after ischemic stroke in animals and humans. J Cereb Blood Flow Metab. 2014;34(2):185–99.
- 121. Chen HS, et al. Glycyrrhizin prevents Hemorrhagic Transformation and improves neurological outcome in ischemic stroke with delayed thrombolysis through targeting peroxynitrite-mediated HMGB1 signaling. Translational Stroke Research. 2020;11(5):967–82.
- 122. The L. 21st century management and prevention of stroke. Lancet. 2018;392(10154):1167.
- Hafez S, et al. Comparative analysis of different methods of ischemia/ reperfusion in hyperglycemic stroke outcomes: interaction with tPA. Trans Stroke Res. 2015;6(3):171–80.
- 124. Bardutzky J, Schwab S. Antiedema therapy in ischemic stroke. Stroke. 2007;38(11):3084–94.
- 125. Ye X, et al. Caspase-1: a promising target for preserving blood-brain barrier integrity in acute stroke. Front Mol Neurosci. 2022;15:856372.
- 126. Van Opdenbosch N, Lamkanfi M. Caspases in cell death, inflammation, and disease. Immunity. 2019;50(6):1352–64.
- 127. Israelov H, et al. Caspase-1 has a critical role in blood-brain barrier injury and its inhibition contributes to multifaceted repair. J Neuroinflammation. 2020;17(1):267.
- 128. Huang Y, et al. Crosstalk between inflammation and the BBB in Stroke. Curr Neuropharmacol. 2020;18(12):1227–36.
- Qiu YM, et al. Immune cells in the BBB disruption after acute ischemic stroke: targets for immune therapy? Front Immunol. 2021;12:678744.
- 130. Maida CD, et al. Neuroinflammatory mechanisms in ischemic stroke: focus on cardioembolic stroke, background, and therapeutic approaches. Int J Mol Sci. 2020;21(18):6454.
- Li Y, et al. New progress in the approaches for blood-brain barrier protection in acute ischemic stroke. Brain Res Bull. 2019;144:46–57.
- 132. Yirong Y, et al. Non-invasive vagus nerve stimulation reduces bloodbrain barrier disruption in a rat model of ischemic stroke. Brain Stimul. 2018;11(4):689–98.
- 133. Li L, et al. The specific role of reactive astrocytes in stroke. Front Cell Neurosci. 2022;16:850866.
- Qu Y, Wang L, Mao Y. Gallic acid attenuates cerebral ischemia/re-perfusion-induced blood-brain barrier injury by modifying polarization of microglia. J Immunotoxicol. 2022;19(1):17–26.
- 135. Zhou X, et al. Mitochondrial dynamics: a potential therapeutic target for ischemic stroke. Front Aging Neurosci. 2021;13:721428.
- 136. Kim Y, et al. Effects of natural polyphenols on oxidative stress-mediated blood-brain barrier dysfunction. Antioxid (Basel). 2022;11(2):197.
- 137. Kadir RRA, Alwjwaj M, Bayraktutan U. Treatment with outgrowth endothelial cells protects cerebral barrier against ischemic injury. Cytotherapy. 2022;24(5):489–99.
- Alwjwaj M, Kadir RRA, Bayraktutan U. Outgrowth endothelial progenitor cells restore cerebral barrier function following ischaemic damage: the impact of NOX2 inhibition. Eur J Neurosci. 2022;55(6):1658–70.
- Zeng X, et al. Zebularine protects against blood-brain-barrier (BBB) disruption through increasing the expression of zona occludens-1 (ZO-1) and vascular endothelial (VE)-cadherin. Bioengineered. 2022;13(2):4441–54.
- Nilles KL, et al. Blood-brain barrier transporters: opportunities for therapeutic development in ischemic stroke. Int J Mol Sci. 2022;23(3):1898.
- 141. Song S, et al. Activation of endothelial Wnt/β-catenin signaling by protective astrocytes repairs BBB damage in ischemic stroke. Prog Neurobiol. 2021;199:101963.
- 142. Martin M, et al. Engineered wnt ligands enable blood-brain barrier repair in neurological disorders. Science. 2022;375(6582):eabm4459.

- Hui J, et al. Fluoxetine regulates neurogenesis in vitro through modulation of GSK-3β/β-catenin signaling. Int J Neuropsychopharmacol. 2015;18(5):pyu099.
- Laksitorini MD, et al. Impact of Wnt/β-catenin signaling on ethanolinduced changes in brain endothelial cell permeability. J Neurochem. 2021;157(4):1118–37.
- 145. Ya-Bin J, et al. Lithium alleviates blood-brain barrier breakdown after cerebral ischemia and reperfusion by upregulating endothelial Wnt/βcatenin signaling in mice. Neuropharmacology. 2021;186:108474.
- 146. Liu C, et al. GLP-1R agonist exendin-4 protects against hemorrhagic transformation induced by rtPA after ischemic stroke via the Wnt/βcatenin signaling pathway. Mol Neurobiol. 2022;59(6):3649–64.
- 147. Huang LY, et al. Healthy serum-derived Exosomes improve neurological outcomes and protect blood-brain barrier by inhibiting endothelial cell apoptosis and reversing autophagy-mediated tight Junction protein reduction in Rat Stroke Model. Front Cell Neurosci. 2022;16:841544.
- 148. Nozohouri S, Vaidya B, Abbruscato TJ. Exosomes in ischemic stroke. Curr Pharm Des. 2020;26(42):5533–45.
- 149. Helms HC, et al. In vitro models of the blood-brain barrier: an overview of commonly used brain endothelial cell culture models and guidelines for their use. J Cereb Blood Flow Metab. 2016;36(5):862–90.
- Claesson-Welsh L, Dejana E, McDonald DM. Permeability of the endothelial barrier: identifying and reconciling controversies. Trends Mol Med. 2021;27(4):314–31.
- 151. Jain KK. Nanobiotechnology-based drug delivery to the central nervous system. Neurodegener Dis. 2007;4(4):287–91.
- 152. Sifat AE, Vaidya B, Abbruscato TJ. Blood-brain barrier protection as a therapeutic strategy for acute ischemic stroke. Aaps j. 2017;19(4):957–72.
- 153. Fukuta T, Oku N, Kogure K. Application and utility of liposomal neuroprotective agents and biomimetic nanoparticles for the treatment of ischemic stroke. Pharmaceutics. 2022;14(2):361.
- 154. Huang Z, et al. A biomimetic zeolite-based nanoenzyme contributes to neuroprotection in the neurovascular unit after ischaemic stroke via efficient removal of zinc and ROS. Acta Biomater. 2022;144:142–56.
- Zhang S, et al. Brain-targeting, acid-responsive antioxidant nanoparticles for stroke treatment and drug delivery. Bioact Mater. 2022;16:57–65.
- Wang D, et al. Urokinase loaded black phosphorus nanosheets for sequential thrombolysis and reactive oxygen species scavenging in ischemic stroke treatment. Biomater Sci. 2022;10(16):4656–66.
- Zhang S, et al. Erythrocyte membrane-enveloped salvianolic acid B nanoparticles attenuate cerebral ischemia-reperfusion Injury. Int J Nanomedicine. 2022;17:3561–77.
- 158. Zhang Y, et al. Nanogels as Novel Nanocarrier Systems for efficient delivery of CNS therapeutics. Front Bioeng Biotechnol. 2022;10:954470.
- 159. Parvez S, et al. Dodging blood brain barrier with nano warriors: novel strategy against ischemic stroke. Theranostics. 2022;12(2):689–719.
- 160. Wang Y, et al. Synergy of Yiqi and Huoxue components of Qishen Yiqi formula in Ischemic stroke protection via lysosomal/inflammatory mechanisms. J Ethnopharmacol. 2022;293:115301.
- Ye Y, et al. Efficacy of chinese herbal medicine for tPA thrombolysis in experimental stroke: a systematic review and meta-analysis. Phytomedicine. 2022;100:154072.
- 162. Zhu QY, et al. Borneol enhances the protective effect against cerebral ischemia/reperfusion injury by promoting the access of astragaloside IV and the components of Panax notoginseng saponins into the brain. Phytomedicine. 2022;94:153822.
- Su XT, et al. Mechanisms of Acupuncture in the Regulation of Oxidative Stress in Treating Ischemic Stroke. Oxid Med Cell Longev. 2020;2020:7875396.
- 164. Kawabori M, et al. Clinical trials of stem cell therapy for cerebral ischemic stroke. Int J Mol Sci. 2020;21(19):7380.
- 165. Boese AC, et al. Neural stem cell therapy for subacute and chronic ischemic stroke. Stem Cell Res Ther. 2018;9(1):154.
- Dabrowska S, et al. Neuroinflammation as a target for treatment of stroke using mesenchymal stem cells and extracellular vesicles. J Neuroinflammation. 2019;16(1):178.
- Xu Y, et al. Exosomal microRNAs as potential biomarkers and therapeutic agents for acute ischemic stroke: new expectations. Front Neurol. 2021;12:747380.

- 168. Ghori A, et al. Vascular endothelial growth factor augments the tolerance towards cerebral stroke by enhancing neurovascular repair mechanism. Transl Stroke Res. 2022;13(5):774–91.
- 169. Daneman R, et al. The mouse blood-brain barrier transcriptome: a new resource for understanding the development and function of brain endothelial cells. PLoS ONE. 2010;5(10):e13741.
- Francisco DMF, et al. Advancing brain barriers RNA sequencing: guidelines from experimental design to publication. Fluids Barriers CNS. 2020;17(1):51.
- 171. He L, et al. Single-cell RNA sequencing of mouse brain and lung vascular and vessel-associated cell types. Sci Data. 2018;5:180160.
- 172. Song HW, et al. Transcriptomic comparison of human and mouse brain microvessels. Sci Rep. 2020;10(1):12358.
- 173. Xie Y, et al. Key molecular alterations in endothelial cells in human glioblastoma uncovered through single-cell RNA sequencing. JCI Insight. 2021;6(15):e150861.
- 174. Zhang Q, et al. Blocking C3d(+)/GFAP(+) A1 astrocyte conversion with Semaglutide attenuates blood-brain barrier disruption in mice after ischemic stroke. Aging Dis. 2022;13(3):943–59.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

