REVIEW Open Access



Identification of the molecular mechanism of insulin-like growth factor-1 (IGF-1): a promising therapeutic target for neurodegenerative diseases associated with metabolic syndrome

Archana Arjunan¹, Dhiraj Kumar Sah^{2,3}, Minna Woo⁴ and Juhyun Song^{1,3*}

Abstract

Neurodegenerative disorders are accompanied by neuronal degeneration and glial dysfunction, resulting in cognitive, psychomotor, and behavioral impairment. Multiple factors including genetic, environmental, metabolic, and oxidant overload contribute to disease progression. Recent evidences suggest that metabolic syndrome is linked to various neurodegenerative diseases. Metabolic syndrome (MetS) is known to be accompanied by symptoms such as hyperglycemia, abdominal obesity, hypertriglyceridemia, and hypertension. Despite advances in knowledge about the pathogenesis of neurodegenerative disorders, effective treatments to combat neurodegenerative disorders caused by MetS have not been developed to date. Insulin growth factor-1 (IGF-1) deficiency has been associated with MetS-related pathologies both in-vivo and in-vitro. IGF-1 is essential for embryonic and adult neurogenesis, neuronal plasticity, neurotropism, angiogenesis, metabolic function, and protein clearance in the brain. Here, we review the evidence for the potential therapeutic effects of IGF-1 in the neurodegeneration related to metabolic syndrome. We elucidate how IGF-1 may be involved in molecular signaling defects that occurs in MetS-related neurodegenerative disorders and highlight the importance of IGF-1 as a potential therapeutic target in MetS-related neurological diseases.

Keywords Alzheimer's disease (AD), Insulin-like growth factor-1 (IGF-1), Metabolic syndrome (MetS), Neurodegeneration, Neuroprotection

*Correspondence:

Juhyun Song

juhyunsong@chonnam.ac.kr

Introduction

Metabolic syndrome (MetS) is a collection of metabolic abnormalities, including hypertension, central obesity, and atherogenic dyslipidemia [1]. MetS significantly increases the risk of type 2 diabetes mellitus (T2DM) and cardiovascular disease [2]. Additionally, emerging evidences have shown that MetS can affect the central nervous system (CNS) diseases through various mechanisms [3]. Several studies suggest that MetS is associated with various neurodegenerative disorders, including



© The Author(s) 2023. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/ficenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

¹ Department of Anatomy, Chonnam National University Medical School, Hwasun, Jeollanam-Do 58128, Republic of Korea

² Department of Biochemistry, Chonnam National University Medical School, Hwasun 58128, Republic of Korea

³ BioMedical Sciences Graduate Program (BMSGP), Chonnam National University, 264 Seoyangro, Hwasun 58128, Republic of Korea

⁴ Division of Endocrinology and Metabolism, University Health Network and and Banting and Best Diabetes Centre, University of Toronto, Toronto, ON, Canada

Arjunan et al. Cell & Bioscience (2023) 13:16 Page 2 of 18

Alzheimer's disease (AD), Huntington's disease (HD), and Parkinson's disease (PD) [4–8].

Synaptic and glial dysfunction with aberrant networks between these cells is a hallmarks of neurodegenerative diseases (NDDs) [9]. Many NDDs can be classified as pyramidal and extrapyramidal, with motor and behavioral or cognitive impairments being the most common clinical manifestations [10]. Various molecular and cellular pathologies are associated with these NDDs, including oxidative stress, mitochondrial dysfunction, calcium (Ca²⁺) influx, glutamate toxicity, proteolytic stress, protein aggregation, neuroinflammation, and neuronal death [11, 12]. Over the past two decades, there has been a significant increase in evidence demonstrating the potent neuroprotective effects of neurotrophic factors (NTFs) on NDDs [13]. NTFs are crucial for CNS development and play vital roles in neurogenesis, neuronal cell migration, and CNS cell survival [14]. Recent research has focused on NTFs to understand their role in the etiology and as potential therapy for various neurological diseases. One of the major NTFs is insulin-like growth factor-1 (IGF-1), a peptide hormone (7649 Da and 70 amino acids) that belongs to the insulin-like hormone superfamily that include insulin, IGF-1, and IGF-2 [15].

The molecular signalling of IGF is highly evolutionarily conserved. IGF1 can act through autocrine, paracrine and endocrine mechanisms to regulate cellular growth, differentiation and proliferation [16]. The IGF system consists of six IGF binding proteins (IGFBPs) and two growth factors (IGF-1 and IGF-2) along with their cognate insulin growth factor receptors (IGF-1R, IGF-2R) [15]. The majority, up to 99% of IGF-1 binds to circulating IGFBP-1[17]. In the brain (hippocampus, cortex, olfactory lobes, cerebellum, and amygdala), IGF-1 binds to IGFBP-2, -4, and -5b [18]. In adults, IGF-1 is produced primarily in the liver and to a lesser extent in the hippocampus, cerebellum, and subventricular zone-olfactory bulb (SVZ-OB) under stimulation of growth hormone (GH) [19]. GH regulates neurogenesis and neuronal plasticity [20]. IGF-1 exerts its actions by binding and activating its membrane receptors, which are receptor tyrosine kinases [16]. After IGF-1 binds to its ligand, a series of phosphorylation events leading to activation of insulin receptor substrates, mitogen-activated protein kinase (MAPK), and phosphoinositide 3-kinase/protein kinase B (PI3K-Akt) lead to various intracellular processes [21]. A recent study has focused on the role and therapeutic potential of IGF-1 in the CNS to improve brain function and complex mechanisms of the CNS in MetS-induced neurodegeneration [22]. Herein, we focus on the potential therapeutic effects of IGF-1 in NDD associated with MetS and the molecular mechanisms underlying its pharmacological effects.

IGF-1 in the CNS

IGF-1 can cross the blood brain barrier (BBB) and enter CSF, and perform a number of important functions of the CNS, including neurogenesis and neuroprotection, through autocrine/paracrine or endocrine effects. It affects metabolic regulation in the CNS, promotion of other nerve growth factors, clearance of aggregate proteins, and angiogenesis [23-25] (Fig. 1). High levels of IGF-1 are found in the CNS during early stages of organogenesis, which promotes brain derived growth factor (BDNF) and other neutrotropic factors that play important roles during brain development [26, 27]. Another study demonstrated that IGF-1 administration increased overall BDNF and decreased expression of interleukin (IL)-1β, TNF-α, nitric oxide synthase (iNOS), and glial fibrillary acidic protein (GFAP) in the whole brain [28]. IGF-1/IGF1R signaling has also been associated with Schwann cell (SC) survival, migration, proliferation, and myelination [29, 30]. In-vitro experiments with glial cells, oligodendrocytes, brain explants, and adult stem cells have revealed that IGF-1 promotes myelination, differentiation and mitogenesis [31]. Furthermore, IGF-1 can promote oligodendroglial cells to survive by inhibiting caspase-3 [31]. IGF-1/IGF-1R knockout mice showed decreased brain size, loss of myelination, and cognitive decline, whereas overexpression of IGF-1 resulted in increased brain size and myelination [32]. Moreover, IGF-1 regulates neural stem cell proliferation by promoting replicative lifespan and shortening all cell cycle lengths, particularly the G1/S transition [33]. Numerous clinical studies have demonstrated that IGF-1/ IGF-1R mutations are associated with mental retardation and microcephaly [34, 35] (Table 2). Lichtenwalner et al., reported that altered levels of IGF-1 negatively affect neurogenesis and synaptic plasticity, particularly in the hippocampus [36]. In an in-vivo models, IGF-1 influences adult dentate gyrus development by increasing the number of granule neurons and thus increasing the dentate granule cell layer [37, 38]. IGF-1-RIT1-Akt-Sox2 pathway plays a key role in IGF-1-induced neurogenesis, cellular proliferation, and gene expression in hippocampus neurons [39]. IGF-1 can also influence neuronal excitability and glutamate system in brain [40]. In various in-vivo and in-vitro studies, exogenous administration of IGF-1 mediated has been shown to increase glucose utilization, release acetylcholine from neurons, activate N-methyl-D-aspartate receptor (NMDA), protect the cerebromicrovascular environment, and maintenance of synaptic structure and function [36, 41-45]. IGF-1 can also interact with NMDA receptors to promote synaptic function and facilitate PI3K/glutamatergic transmission in the hippocampus [46–49]. Kelsch et al. showed that during hippocampal maturation, K+/Cl- outward

Arjunan et al. Cell & Bioscience (2023) 13:16 Page 3 of 18

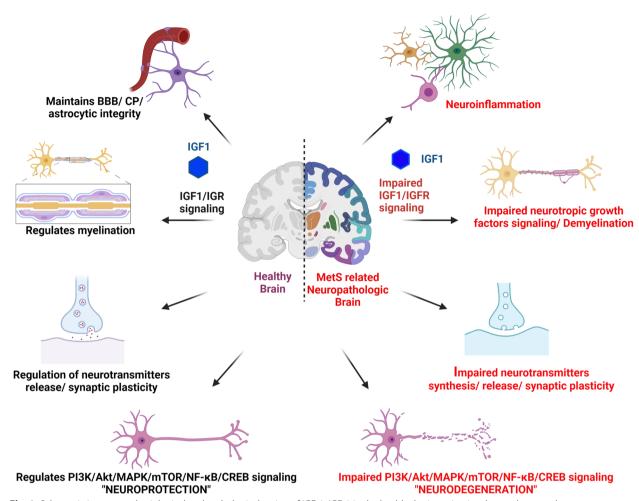


Fig. 1 Schematic image on physiological and pathological action of IGF-1. IGF-1 in the healthy brain maintains the cerebrovascular microenvironment and BBB/CP integrity, regulates inflammation in microglia, and facilitates synaptic communication and cognition by acting on ionic channels and neurotransmitters. In Metabolic syndrome (MetS) related brain, both systemic and local deficiency of IGF-1 shows the altered cerebrovascular microenvironment/disturbed BBB/CP integrity, increased the deposition of α- Synuclein/Tau/Aβ/HTT proteins cause impaired neuroinflammatory action, neurotransmitter release, synaptic plasticity, and cognition which may lead to neurodegenerative diseases (AD/PD/HD)

transport is mediated by IGF-1/PI3K pathway [50]. Furthermore, IGF-1 increased presynaptic facilitation by activating p38/MAPK to modulate K⁺ channel activity [51, 52]. However, IGF-1R is highly expressed in cerebral plexus (CP), hypothalamus, thalamus, amygdala, and hippocampus/parahippocampal gyrus. Given that these regions are critically linked to cognition, it is compelling that IGF-1 and IGF-1R deficiencies lead to cognitive impairment [46, 53]. The transcriptional regulator CREB (cAMP response element binding protein) is a critical regulator of axonal growth and neuronal plasticity that is important for neuroprotection and cognition preservation [54]. In many cell types, IGF-1 enhances CREB phosphorylation and controls CRE-containing genes, such as c-Fos and B-Cell Leukemia/Lymphoma 2 (Bcl-2)

[55]. Neuronal survival is also linked to the MAPK-CREB signaling pathway. By phosphorylating Bad and CREB, activated ribosomal protein S6 kinase beta (RSKs) can inhibit apoptosis [56, 57]. Also, IGF-1 can suppress various proapoptotic signals through regulation of multiple downstream targets [55]. As such, IGF-1 is an essential factor in maintaining neuronal homeostasis, and identifying the role of IGF-1 in the brain is important for finding clues to effective treatment targets for NDDs (Fig. 1).

Therapeutic applications of IGF-1 in neurological diseases

Several recent studies highlight the pleiotropic actions of IGF-1 in neurons [29, 39, 42]. Tables 1, 2, and 3 describe the consequences of IGF-1 deficiency and the therapeutic

Arjunan et al. Cell & Bioscience (2023) 13:16 Page 4 of 18

 Table 1
 IGF-1-deficient-induced neurological disease

| No | Model | Findings | References |
|----|--|---|------------|
| 1 | LID Mice | IGF-1 deficient cause neuro-glio-vascular unit damage | [188] |
| 2 | IGF1R (VE-Cadherin-Cre ^{ERT2} /lgf1r ^{f/f}) | IGF-1 is critical for cerebromicrovascular endothelial health and maintenance of normal neurovascular coupling (NVC) responses | [189] |
| 3 | IGF1R (GFAP-Cre ^{ERT2} /lgf1r ^{f/f}) | IGF-1 promotes astrocyte health and maintains normal NVC, protecting cognitive health | [45] |
| 4 | AD clinical study | The IGF-1 level was increased in AD subjects' serum but not in CSF | [190] |
| 5 | Aged LID mice | IGF-1 is essential for the regulation of mitochondrial function, redox status, and cognition, and IGF-1 deficiency with age may increase brain damage and cognitive deficits | [191] |
| 6 | igfr ^{f/f} mice | Reduced IGF-1 increases the accumulation of extrasynaptic glu- tamate, which may contribute to neurodegeneration in disease states | [192] |
| 7 | IGFR (GFAP-Cre ^{TAM} /igfr ^{f/f}) | Reduction in IGFR expression with age is associated with a decrease in hippocampal-dependent learning and increased gliosis | [193] |
| 8 | MS clinical study | Low serum IGF-1 was associated with cognitive impairment and fatigue in MS | [194] |
| 9 | AD and vascular dementia (VaD) clinical study | Low serum IGF-1 was a risk marker for VaD | [195] |
| 10 | AD clinical study | Lower baseline serum IGF-1 was associated with a faster cognitive decline in AD over a 2-year period | [196] |
| 11 | IGF-1 deficient mice (Igf1 ^{f/f} +TBG-Cre-AAV8) | IGF-1 deficiency exerts deleterious effects on cerebral microcirculation, causes a decline in cortical and hippocampal capillarity, and exacerbates hypertension-induced cerebromicrovascular rarefaction | [42] |
| 12 | AD clinical study | Increased levels of circulating IGF-1 and IGFBP-3 cause differences in mean age and MMSE scores, and circulating levels of IGFBP-3 decrease the level of IGF-1 | [197] |
| 13 | AD clinical study | Lower serum IGF-1 was associated with cognitive impairment and was involved in the pathogenesis of cognitive deficits in AD | [84] |
| 14 | Postnatal/adult global IGF-I knockout (KO) mice (Igf-I2/2) | IGF-1 regulates postnatal/adult hippocampal neurogenesis in a stage-dependent manner | [198] |
| 15 | IGF-1 deficiency (Igf1(f/f)-TBG-Cre-AAV8) | IGF-1 deficiency also impaired glutamate-mediated CBF responses, likely due to dysregulation of astrocytic expression of metabotropic glutamate receptors and impaired mediation of CBF responses by eicosanoid gliotransmitters | [169] |
| 16 | IGF-1 deficiency (Igf1(f/f) -TBG-Cre-AAV8) | IGF-1-deficient mice included exacerbated disruption of the BBB and neuroinflammation that were associated with impaired hippocampal cognitive function | [164] |
| 17 | Prenatal stress/Dawley/Adult male offspring IGF-1, 10–20 μ g/h/i.c.v | IGF-1 administration decreased IGF-1 levels and IGF-1 phosphorylation with altered IRS-1 phosphorylation in the hippocampus and frontal cortex of prenatal stress-induced rats | [199] |
| 18 | AD clinical study | Reduced serum IGF-1 is associated with the development of AD dementia in patients with AD | [82] |
| 19 | Clinical study IGF-1R mutation | IGF-1R mutations lead to prenatal and postnatal growth retardation and microcephaly | [27] |
| 20 | AD clinical study | Patients with AD as well as other dementias had high levels of IGF-1 in serum but not in CSF | [200] |
| 21 | AD clinical study | Low serum levels of IGF-1 and IGFBP-3 in males with AD but not in females with AD $$ | [201] |
| 22 | Viral-mediated Cre-lox P system to knockout the lgf1 gene animal model | Adult-onset IGF-1 deficiency alone is sufficient to induce a depressive phenotype in mice Individuals with low brain IGF-1 levels are at increased risk for depression, and these behavioral effects are not ameliorated by increased local IGF-1 production or transport | [202] |

Arjunan et al. Cell & Bioscience (2023) 13:16 Page 5 of 18

Table 1 (continued)

| No | Model | Findings | References |
|----|---|--|------------|
| 23 | AD clinical study | Significant decreases in IRS-1 and IRS-2 levels were identified in AD neurons in association with increased levels of inactivated phosphor (Ser312) IRS-1 and phosphor(Ser616)IRS-1, where increased levels of these phosphoserine epitopes colocalized strongly with NFTs | [203] |
| 24 | APP (SW), Tg2576 mice | Impaired IGF-1/IRS-2 signaling prevents premature death and delays amyloid accumulation in a model of AD | [204] |
| 25 | IGF-1R [±] mice/MPTP induction | IGF-1R $^\pm$ mice have shown increased dopamine neuronal loss in MPTP-induced mice | [96] |
| 26 | AD clinical study | Patients with vascular dementia and AD had low IGF-1 that may cause carotid atherosclerosis | [205] |
| 27 | Clinical cohort study IGF-1 and IGF-1R mutation | IGF-1 and IGF-1R mutant children had intrauterine growth retardation and poor postnatal growth | [26] |
| 28 | Brain injury-induced Romney-Suffolk fetal sheep/IGF-1 (3 or 30 μ g/i.c.v) | IGF-1 treatment reduced caspase-3 activation and increased glial proliferation in a dose-dependent manner | [31] |

effects of IGF-1 in experimental and clinical studies of neurological diseases. Our focus in this section is the effect of IGF-1 on neurodegenerative diseases, specifically AD and PD.

Alzheimer's disease (AD)

AD is characterized by a progressive cognitive decline affecting around 25 million individuals worldwide [58], and causes difficulties in learning and memory, language, and executive motor function [59]. AD is generally thought to be caused by amyloid-beta (A β) accumulation and plaque formation in the brain, a pathology known as the "amyloid hypothesis" [60]. There are evidences that IGF-1 may prevent age-related cognitive decline [48]. Several studies have shown that low IGF-1 levels are associated with AD (Table 1).

Along with the canonical trophic role of IGF-1, it has also been shown to exert neuromodulatory effects through regulating neurotransmitter release (Fig. 2). Emerging research shows that glutaminergic neurotransmission through glutamate receptor NMDA plays a major role in learning and memory [61-63]. NMDA is also involved in the induction of long term potentiation (LTP) [63, 64] and can regulate synaptic plasticity [65]. Sonntag et al. reported that chronic administration of IGF-1 increases the density of NMDA receptors (NMDAR1, NMDAR2A, and R2B subunits) in the hippocampus, dentate gyrus and cortical areas, which are mainly involved in learning and memory [48]. Trejo et al. showed that IGF-1 restores cognitive function by attenuating the deposition of $A\beta$ in an experimental model of AD [25].

Furthermore, IGF-1 has been implicated in several ways to affect synaptic plasticity [66, 67]. IGF-1 may promote synaptic plasticity and transmission in minutes

or persist for several hours to increase neuronal differentiation and survival (Fig. 2). Several types of neurons become more excitable in response to IGF-1 [68]. Studies have shown that the systemic administration of IGF-1 improves synaptic complexity and neurogenesis in the hippocampus [36, 69]. Moreover, IGF-1 in cultured hippocampal neurons increased the frequency of spontaneous excitatory postsynaptic currents (sEPSCs) for a short or long term, but had no effect on miniature excitatory postsynaptic currents (mEPSCs) or spontaneous inhibitory postsynaptic currents (sIPSCs). Indeed, the excitatory transmissions has been shown to be mediated by MAPK pathways [68]. Furthermore, IGF-1 inhibited synaptic transmission by increasing the frequency of sIPSCs in response to A β - reduction in sIPSC frequency [70]. This suggests that IGF-1 increased glutamate release at presynaptic sites or the functional excitability of synaptic contacts, but had no effect on non-NMDA or NMDA receptors.

In the CA1 region of the hippocampus, des (1-3)-IGF-1 increased the field excitatory postsynaptic potentials (fEPSPs), EPSCs, and α-amino-3-hydroxy-5-methyl-4isoxazolepropionic acid receptor (AMPAR)-mediated postsynaptic exocytosis/endocytosis mechanism [71]. Des-IGF-1 affects glutamate receptor AMPAR binding protein (GRIP), N-ethylmaleimide-sensitive fusion protein (NSF), stargazin, and proteins that interact with C-kinase-1, which influence AMPA receptor anchoring, surface translocation, and synaptic targeting [72]. Furthermore, activation of the IGF-1R facilitates the AMPA synaptic mechanism by increasing intracellular calcium mobilization at the synapse [67]. These findings suggest that IGF-1 is important for regulating the AMPA receptors involved in LTP and cognition. IGF-1 modulates synaptic plasticity primarily by regulating ion channels Arjunan et al. Cell & Bioscience (2023) 13:16 Page 6 of 18

Table 2 Therapeutic applications of IGF-1 in neurological diseases

| No | Model | Findings | References |
|----|---|---|------------|
| 1 | In vivo⁄in vitro AD model | In vivo transduction with RAd-IGF1 blocked memory impairment | [206] |
| 2 | Brain-specific IGF-1 overexpression mice | IGF-1 treatment reduced depressive and anxiety-like behavior, improved motor coordination, motor learning, visuospatial, and working memory | [207] |
| 3 | C57BL/6 J mice/controlled cortical impact/IGF-1 | Increased immature neuronal density and neurogenesis of the hippocampus | [208] |
| 4 | RIT1 ^{-/-} mice/IGF-1 | IGF-1 facilitates hippocampal neurogenesis through the RIT1/Akt/Sox2 signaling pathway | [39] |
| 5 | Old male rats/IGF-1 | IGF-1 increases hippocampal neurogenesis and memory accuracy in aged individuals | [209] |
| 6 | Old Sprague–Dawley female rats/IGF-1, 18 days ICV | IGF-1 treatment increased the branching of hippocampal astrocytes and reduced their number in the hippocampal striatum radiatum, and improved spatial memory accuracy in aging rats | [210] |
| 7 | Female Sprague–Dawley rats/MCAo/IGF-1 | IGF-1 reduced infarct volume (39%) and BBB permeability and suppressed IL-6, IL-10, and TNF- $\!\alpha$ | [211] |
| 8 | SH-SY5Y cells/10 nM IGF-1 | IGF-1-induced shedding of both APP and APLP1 depends on PI3K, while APLP2 shedding is independent of this signaling pathway | [212] |
| 9 | In vitro/PD/IGF-1 along with MPP+ | IGF-1 increases cell viability and decreases cell apoptosis | [213] |
| 10 | SH-EP1 cell lines/IGF-1 MPP ⁺ neurotoxicity | Inhibition of MPP+-induced apoptosis by activating JNK by PI3K/AKT/GSK3 β pathway | [106] |
| 11 | PD (WT, A30P and A53 T mutant)/100 ng/mL | Rescue from $\alpha\text{-synuclein}$ toxicity and suppression of $\alpha\text{-synuclein}$ aggregation | [214] |
| 12 | MT-IGF mice | Inhibits $\beta\mbox{-cell}$ apoptosis, insulin secretion, and hepatic glucose production | [114] |
| 13 | Rat/6-OHDA/IGF-1 transgenic neurospheres | Reduction in amphetamine-induced rotation and increased survival of human neural progenitor cells (hNPC) exert trophic effects on degenerate dopamine neurons in the PD model | [103] |
| 14 | Adult female Long-Evans rats/6-OHDA/MPTP/IGF-1 | By activating PI3K/Akt signaling, IGF-1 improved motor behavior and reduced DA loss in SNc | [105] |
| 15 | APP/PS2 mice IGF-1 (50 g/kg dose, i.p.) | Reverses spatial learning and memory impairment and reduces total brain $\mbox{\rm A}\beta$ deposition | [24] |
| 16 | Male Wistar rats/6-OHDA/GPE (3 mg/kg, i.p.) | Increased motor movement and reduced dopamine neuronal loss in PD rats | [101] |
| 17 | Adult female Long-Evans rats/6-OHDA/MPTP/IGF-1 | IGF-1 significantly reduced the loss of asymmetric movement of the forelimb, reduced SNc neuronal loss, and TH immunoreactivity in DA fibers and striatum | [102] |
| 18 | Wistar rats/LID mice IGF-1: 50 μg/kg/rat/day | Reduced the brain $A\beta$ burden and upregulated the brain levels of $A\beta$ carriers | [83] |
| 19 | In vitro/IGF-1 (0.5 mg/mL)/dopamine | Decrease in apoptosis was accompanied by an increase in Bcl-2 levels | [4] |
| 20 | APP (WT-APP and V642I-APP mutant)/IGF-1: 10 nM | IGF-1 protected cells from APP-induced apoptosis and suppressed the cleavage of procaspase-3 | [5] |
| 21 | Male BN \times F344 rats/IGF-1 (50 ng/0.5 μ L/h, i.c.v) | IGF-1 administration restored neurogenesis via a three-fold increase in neuronal production | [36] |
| 22 | Sprague–Dawley/hx rats/carotid artery IGF-1 infusion; 1.25 mg/kg per day) | IGF-1 increases progenitor cell proliferation and selectively induces neurogenesis in the progeny of adult neural progenitor cells in the hippocampus | [37] |
| 23 | Male Wistar rats/6-OHDA/GPE | A single dose of GPE increased TH immunoreactivity and reduced TH immunoreactive neuronal cell death in SNc and striatum | [100] |

 $(Ca^{2+}\text{-binding}\ proteins),\ neurotransmitter\ secretion,\ and\ neuronal\ arborization. Also, IGF-1\ phosphorylates\ and\ activates\ the\ \alpha-1\ subunit\ of\ the\ L-type\ Ca^{2+}\ channel\ through\ the\ PI3K\ pathway\ [73].\ The\ Na^+/Ca^{2+}\ exchanger\ (NCX)\ is\ a\ neuronal\ reciprocal\ Ca^{2+}\ transporter\ that\ promotes\ neuroprotection. This is\ mediated\ by\ IGF-1\ by\ increasing\ the\ NCX-induced\ inward\ and\ decreasing\ the\ outward\ current\ [74].\ Accordingly,\ systemic\ IGF-1\ modulates\ the\ electrophysiological\ properties\ of\ target\ neurons.\ IGF-1\ blocks\ transient\ A-type\ K^+\ currents\ and\ increases\ high-voltage-activated\ Ca^{2+}\ currents,\ while\ keeping\ low-voltage-activated\ Ca^{2+}\ and\ Na^+\ current\ constant\ [75]\ .$

Other glutamate receptors, including kainate receptors (KARs) and metabotropic glutamate receptors, can control long-term and short-term synaptic plasticity [76, 77]. KARs can be found on the gamma-aminobutyric acid (GABAergic) and glutamatergic presynaptic terminals [78]. IGF-1 increases the potency of kainate-dependent currents in cerebellar granule neurons and modulates Ca²⁺, Cl⁻, and K⁺ channels by PI3K-dependent pathway, but not MAPK dependent pathway [73, 79]. Although IGF-1 can stimulate neurogenesis and promote cognition in short-term, some studies have demonstrated that chronic administration of IGF-1 causes side effects such

Arjunan et al. Cell & Bioscience (2023) 13:16 Page 7 of 18

Table 3 Therapeutic action of IGF-1 on MetS-related neurological diseases

| No | Model | Findings | References |
|----|---|--|------------|
| 1 | C57BL/6J mice/HFD/PEG-IGF-1 | IGF-1 reduced anxiety-like depressive behavior and improved mitochondrial function via CREB/PGC-1α pathway | [183] |
| 2 | C57BL6/J mice/HFD LID mice (subcutaneous hIGF-1, 5 μ g/kg/day) | IGF-1 increased sAPP α /sAPP β ratio, increased peripheral A β clearance | [6] |
| 3 | Male C57BL/6J rats/HFD/HT22 cell line IGF-1, 1 mg/kg/4 weeks | IGF-1 enhanced cognition in HFD rats and inhibited inflammation and oxidative stress in the hippocampus through the activation of the PI3K/Akt/CREB pathway | [7] |
| 4 | Male Sprague–Dawley rats Adult Zucker diabetic fatty (ZDF) rats C57BL/6J mice hIGF-1, 20 μ g/IP | IGF-1 increased CEBP β overexpression, promoted neurite outgrowth, and mitochondrial respiration in diabetic animal models | [29] |
| 5 | Male Sprague–Dawley rats/STZ/IGF-1, 20 μg/subcutaneously | IGF-1 activates and upregulates AMPK to improve mitochondrial function, ATP synthesis, mtDNA copies, and ETS expression levels | [215] |
| 6 | R6/2 mice/IGF-1 | Continuous peripheral administration of IGF-1 partially recovers plasma IGF-1 levels, inhibits HD-related glucose intolerance, protects from weight loss, and improves paw clasping scores | [8] |
| 7 | Male C57BL/6 N mice/STZ IGF1-AAV | IGF-1 improved motor function and reduced muscle atrophy and demyelination of the peripheral motor nerve fibers | [128] |
| 8 | STZ/MCAo model/IGF-1 | Decreased lesion volume (CA1 and CA3 regions of the hip- pocampus and cortex) and reduced apoptosis | [216] |
| 9 | Sprague–Dawley rats/STZ/IGF-1 | Prevented alteration of coenzymes ($\rm Q_9$ and $\rm Q_{10}$) and improved the antioxidant mechanism in diabetes-induced rat's brain, liver, and kidney | [217] |
| 10 | IGF-1 Tg mice bred with IRS-1 null mutant (IRS-1 $^{-/-}$) | IGF-1 overexpression increased brain weight (43%) and promoted oligodendrocyte development and myelination | [130] |
| 11 | Male Sprague–Dawley rats/STZ/sc cell line-10 nM IGF-1 | SCs are effectively protected against glucose-induced apoptosis by IGF-1 | [218] |

as accelerated aging, cancer development, and decreased lifespan [80, 81]. Thus, fine tuning these potential hormonal effects remains an important challenge to addressed.

Researchers have reported that reduced serum IGF-1 levels are associated with AD and decreased brain volume in clinical studies [82]. A study by Carro et al. showed that systemic administration of IGF-1 to mice deficient in hepatic IGF-1 resulted in increased serum IGF-1 levels, decreased the Aß bodies in brain, and increased uptake by Aβ bodies in CSF facilitated by transthyretin and albumin [83]. Kimoto et al. also reported that reduced IGF-1 in serum is associated with cognitive deficits in subjects with AD [84]. IGF-1 can prevent AD development by altering several signaling proteins including rat sarcoma virus (Ras), forkhead box O (FoxO), and MAPK and their pleiotropic actions [85]. The IGF-1R belongs to the tyrosine receptor kinase family that controls many downstream targets, notably MAPK, Akt, Ras, PI3K, and the binding proteins growth factor receptor-bound protein 2 (Grb2) and Shc (Src homology 2 domain containing) transforming protein 1) [86]. PI3K/Akt is a well-known cascade induced by stimulation of IGF-1R [87]. Activated PI3K phosphorylates PIP2 to PIP3, which triggers the phosphorylation of P3-dependent kinase-1/2 (PDK-1/2) at Thr308 and Ser473 residues, resulting in Akt to be recruited to the plasma membrane (Fig. 2). The activated Akt can in turn phosphorylate various target proteins involved in survival and differentiation pathways, including BCL2 associated agonist of cell death (Bad), GSk3, nuclear factor-κB (NF-κB), FoxO1, FoxO3a, and FoxO4 [88]. IGF-1 suppressed NF-κB signalling by upregulating miR-219a-2-3p and inhibiting *YY1* gene expression, which is important for the activation of NF-κB signalling [89].

In AD, elevated levels of tumor necrosis factor- α (TNF- α) may play a significant role in exacerbating amyloidosis [90, 91], and IGF-1 attenuate amyloidosis by antagonizing TNF- α [83]. Recent researches have showed that altered CP function can exacerbate A β accumulation in the brain [73, 92], and numerous in-vitro studies have shown that IGF-1 can maintain tight junction stability in CP epithelial cells [92, 93]. On *in-vitro* study have also shown that IGF-1 maintains tight junction stability in CP epithelial cells [93]. Therefore, IGF-1 can modulate various ion channels and molecular signlaing pathways to attenuate inflammation and promote BBB-CP stability to prevent A β deposition and cognitive decline in AD.

Parkinson's disease (PD)

PD is the second common neurological diseases after AD, with a high incidence among adults in their 50s and 60s [94]. The neuropathological hallmarks of PD are a

Arjunan et al. Cell & Bioscience (2023) 13:16 Page 8 of 18

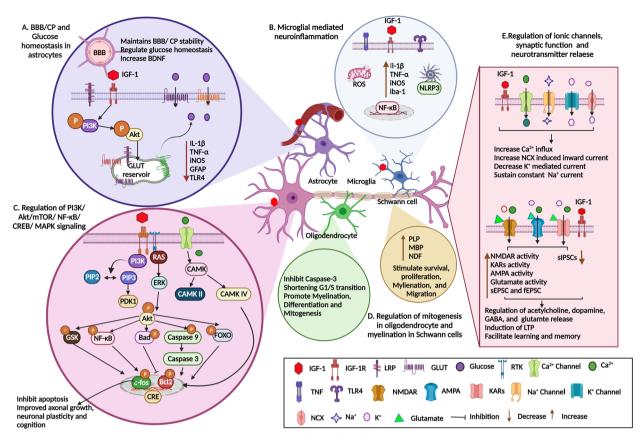


Fig. 2 Schematic image on molecular actions of IGF-1 in CNS cells. **A** Blood–brain barrier (BBB)/choroid plexus (CP) and glucose homeostasis in astrocytes: IGF-1 binds to the astrocytic cell membrane's IGF-1 receptors, activates the PI3K/Akt pathway, and recruits the GLUT transporters, which then begins the uptake of glucose into the cell via GLUT transporters. **B** Neuroinflammation caused by microglia: When IGF-1 binds, it stimulates the polarization of the macrophages via TLR4 increasing the production of IL-1β, TNF-α, iNOS, and iba-1 while decreasing ROS and activating NF-κB/NLRP3 signaling. **C** PI3K/Akt/mTOR/NF-κB/CREB/MAPK signaling regulation in neurons: The PI3K/Akt signaling cascades are initiated when IGF- binds, phosphorylating the GSK, NF-κB, Bad, Caspase 9, and FOXO proteins. These additional phosphorylation result in the nuclear phosphorylation of c-fos and Bcl2, which prevents apoptosis, promotes axon development, and enhances neural plasticity. **D** Regulation of mitogenesis in oligodendrocytes and myelination in Schwann cells: In oligodendrocytes, IGF-1 inhibits the caspase-2 activity, shortening the G1/S cell cycle transition. In Schwann cells, IGF-1 facilitates myelination via increasing the myelinated proteins such as PLP, MBP, and NDF. **E** Regulation of ionic channels, synaptic function, and neurotransmitter release: IGF-1 regulates the Na⁺/Ca²⁺/K⁺ channels to increase the Ca²⁺ influx and maintain the Na⁺ concentration. In neurotransmitters, IGF-1 activates the NMDAR/KAR/AMPA receptors which regulate the acetylcholine, GABA, glutamate, and dopamine synthesis and release

neuronal cell damage in substantia nigra of brain, leading to insufficient secretion of dopamine and accumulation of intracellular inclusions including α -synuclein aggregates [95]. Effective treatment for individuals with PD is challenging due to the lack of pharmacological options and adverse effects such as dyskinesia related to the use of levodopa.

In 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP)-induced mouse model of PD, IGF-1R deficiency resulted in enhanced dopaminergic neuronal death [96]. Several clinical studies showed that low serum IGF-1 level was present in individuals with PD [97–99]. A cleaved form of IGF-1, glycine-proline-glutamate (GPE), prevents the death of tyrosine hydroxylase (TH)

immunopositive neurons, and restores TH immunore-activity in the substantia nigra compacta (SNc) and the striatum of a 6-hydroxydopamine (6-OHDA)-induced PD model [100].

Administration of cleaved IGF-1 (GPE-3 mg/kg, intraperitoneally [i.p]) also improved motor function and decreased dopaminergic neuronal loss in the 6-OHDA model [101]. Similarly, treatment with IGF-1 in 6-OHDA-induced PD model of ovariectomized rats resulted in increased motor function of the forelimbs, reduced loss of SNc neurons, and normal immunoreactivity of TH in the striatum and dopaminergic fibers [102]. In another study, IGF-1 significantly upregulated the survival of human neural progenitor cells in the

Arjunan et al. Cell & Bioscience (2023) 13:16 Page 9 of 18

6-OHDA-induced PD model [103]. Alessandro et al. found that after depolarization, dopaminergic neurons secrete IGF-1, which can stimulate dopamine release in the ventral midbrain [104]. The neuroprotective effects of IGF-1 on PD are mediated by PI3K/Akt signaling rather than MAPK/ERK pathway [105]. Wang et al. reported that IGF-1 inhibits the activation of c-Jun N-terminal kinases (JNK) via the PI3K/AKT/GSK3 β (Glycogen synthase kinase 3 β) pathway, and 1-methyl-4-phenylpyridine ion (MPP⁺)-induced apoptosis [106]. Therefore, IGF-1 prevents the loss of dopaminergic neurons and improves motor function in PD model by upregulating the PI3K/AKT/GSK3/MAPK/ERK pathway.

Metabolic syndrome and neuropathology

MetS is observed concurrently with several abnormalities including central obesity, hyperglycemia, hypertension, dyslipidemia, inflammation and thrombotic states [60]. The International Diabetes Federation criteria for MetS included a fasting blood glucose levels > 5.6 mmol/L (100 mg/dL); blood pressure > 130/85 mmHg; blood triglyceride levels > 1.7 mmol/L (150 mg/dL); HDL cholesterol levels < 1.0 mmol/L (40 mg/dL) for men and < 1.3 mmol/L (50 mg/dL) for women, and waist circumference > 94 cm (men) or > 80 cm (women) [107, 108]. Considering recent evidences, MetS is a major risk factor for type 2 diabetes (T2D) and cardiovascular disease, as well as an emerging major risk factor for NDDs.

Accumulating evidence supports that MetS plays a major role in the development of cognitive impairment [109]. MetS is also known to induce oxidative stress and inflammation, which can lead to cognitive decline by reducing the number and function of hippocampal neurons [109–112]. Furthermore, studies investigated the relationship between circulating IGF-1 concentrations and metabolic syndrome. This review focused on the neuroprotective effects of IGF-1 in MetS-related NDDs.

Diabetes mellitus-related neurodegenerative disease

Diabetes mellitus is characterized by hyperglycemia due to complex pathogenic mechanisms involving widespread insulin resistance and impaired insulin production. Type 1 diabetes (T1D) is an autoimmune disease that causes damage to pancreatic β -cells. The most common type, type 2 diabetes (T2DM), is characterized by dysfunctional β cells and insulin resistance [113]. Scientific evidences has demonstrated a substantial association between diabetes (both T1D and T2D) and cognitive decline leading to dementia in animal models and humans [29, 49, 114, 115].

One study shows that 56% of AD dementia area associated with T2D [116]. In fact, the significance of the

link between T2D and AD is now defined by the term "type 3 diabetes", which describes a subset of diabetic patients who develop AD dementia [117-119]. In T2D, insulin resistance and altered IGF-1/IGF-1R signaling are associated with cognitive decline, AB production, tau hyperphosphorylation, proinflammatory marker's expression, oxidative stress, and dyslipidemia [120, 121]. Rui-Hua et al. found that decreased serum IGF-1 levels were associated with T2D-associated cognitive decline in clinical trials [122]. Another study showed that subjects with mild cognitive impairment with T2D had a reduced serum IGF-1/IGFBP-3 molar ratio [123]. Aksu et al. showed that reduced IGF-1 induces anxiety-like behavior and reduced blood flow to the prefrontal cortex in streptozotocin (STZ)-induced diabetic rats [124]. In addition, Jing et al. showed that maternal hyperglycemia reduces the expression of IGF-1, resulting in delayed fetal dendrite development in STZ-induced rats [125].

Hyperglycemia is associated with a lack of neurotrophic signaling that can lead to mitochondrial dysfunction of SC [126]. Chronic hyperlgycemia can lead to vacuolization and atrophy or degeneration of myelinated nerve fibers [127]. Myelinated nerve fibers (A δ -type afferent fibers) are susceptible to dysfunction when their conduction velocity changes [127]. Chu et al. reported that STZinduced mice carrying an IGF-1 adeno-associated viral (AAV) vector showed reduced peripheral motor nerve fiber demyelination [128]. SC express IGF-1 receptor, and activation by IGF-1 stimulates myelination, attachment to axons, and migration [129]. Ping et al. found that in the cerebral cortex and brainstem, IGF-1 increased the expression of proteins essential for myelination, such as the proteolipid protein (PLP) and myelin basic protein (MBP)[130]. IGF-1 promotes Po induction, DMA synthesis, and DNA synthesis caused by neuro-differentiation factor isoforms in SC. These findings demonstrate that IGF can stimulate proliferation and differentiation in SCs [128].

IGF-1 forms the central core elements of astrocyte functions, such as the regulation of glucose uptake, glutamate transport, and protection against oxidative stress in the brain [44, 131, 132]. IGF-1R enters astrocytes by binding to astrocyte glucose transporter 1 (GLUT1) via the low-density lipoprotein receptor-related protein-1 (LRP1) and scaffolding protein GIPC PDZ domain containing family, member 1 (GIPC1) (Fig. 2). These results suggest that IGF-1R modulates brain glucose metabolism by inhibiting the activity of GLUT1 in astrocytes [44]. Another study demonstrated that IGF-1 increased hypoxia-inducible factor-1 (HIF-1) and GLUT3 protein expression to maintain glucose homeostasis in neurons through PI3K/Akt/mTOR-dependent pathway [133]. These results imply that astrocytes may be important

Arjunan et al. Cell & Bioscience (2023) 13:16 Page 10 of 18

sensors of peripheral hormonal changes that connect the cerebral microenvironment to neurons to respond to endocrine signals. Therefore, therapeutic targets for improving astrocytic function include enhancement of IGFR signalling and mitochondrial function and glucose transport, which can alleviate age-related pathologies such as AD (Fig. 2).

On the other hand, in diabetic mice, IGF-1 expression was significantly decreased and pain, neuroinflammation, and M1 microglial polarization were increased [134]. Microglia are highly dynamic and can adopt wideranging responses to their environment to govern CNS homeostasis [135]. In brain injury, the microglia response switches from a proinflammatory M1 to an anti-inflammatory/reparative M2 for recovery. If this process is not regulated, excessive reactive nitrogen species (RNS), ROS, and inflammatory cytokines secreted by M1 phenotype microglia can cause neuronal damage [135–137]. IGF-1 is mainly produced by microglia, which is elevated during the inflammatory process [138, 139]. IGF1 as a pleiotropic hormone, signals macrophages to help various tissues develop and maintain homeostasis [140]. Sun et al. reported that IGF1R stimulates M1 polarization through toll-like receptor (TLR4)/NF-κB pathway in intracerebral haemorrhage (ICH) induced mice [141]. Furthermore, IGF-1 activates the PI3K/Akt/FoxO1 pathway without affecting TLR2/4 expression in an in vitro hyperglycemic study [142]. Wolters et al. demonstrated that IGF-1 does not produce cytokine itself, and regulates TLRs responsible for inflammatory effects during metabolic complications. Another study reported that TLR4 mutant mice fed HFD showed neurovascular protection by improving astrocytic vascular recovery and cerebromicroenvironement [143]. Similarly, Maria et al. reported the anti-inflammatory action of IGF-1 in astrocytes by IGF-1 gene therapy. Additionally, exogenous treatment with IGF-1 reduced TLR4 expression and reduced NF-κB activation in lipopolysaccharide-induced inflammatory response of astrocytes [144]. As a result of TLR activation, downstream signaling pathways such as PI3K/Akt/ mTOR and MAPK are induced, and promote cytokines production through activation of the NF-κB signaling pathway. These downstream targets are shared by the IGF1 receptor and TLRs [145]. Lee et al. demonstrated that IGF-1 exerts anti-inflammatory action by downregulating the TLR4 signaling in skeletal muscle [146]. These findings suggest direct pro/anti-inflammatory actions of IGF-1 which regulates neuroinflammation and is involved in neuroprotection by maintaining the cerebromicroenvirontment, increasing the capillary density and microglial activation in neuroinflammation by TLR4 signaling (Fig. 2). These findings imply that decreased IGF-1 levels are directly related to cognitive impairment,

and neuroinflammation, and suggest that therapeutic restoration of IGF-1 levels may improve cognitive function.

Obesity-related neuropathology

The prevalence of MetS has increased dramatically in the past decades, primarily due to significant lifestyle changes, including imbalance diet and physical inactivity [147]. According to recent estimates, around 2.1 billion people are overweight or obese [148]. Obesity has become a global epidemic with enormous medical, social, and economic burdens. Western diets are high in salt, processed carbohydrates and saturated fats, which negatively impact body mass and metabolism, including dyslipidemia, abdominal obesity, and T2D [149].

Obesity negatively affects CNS homeostasis and cognitive function [148, 150, 151]. The CNS and peripheral nervous system are fundamentally different in structure and function. And since both are prone to obesity-related dysfunction, this suggests a common pathway leading to the persistent disease progression through visceral fat. Also, a high body mass index (BMI) (>30 kg/m²) has been recognized as one of the risk factors for PD [152, 153]. Obese individuals have fewer striatal dopamine receptors than non-obese individuals. Obesity has a deleterious influence on motor function and manual dexterity [154, 155].

Additionally, Bhat et al., reported that a high-fat/highcholesterol diet can promote cognitive decline and brain dysfunction [156]. High fat diet (HFD)-induced obesity altered the circulating IGF cascade and increased circulatory level of total IGF-1, IGF-2, free IGF-1, and IGFBP3 in rodent and clinical trials [157, 158]. However, insulin/ IGF signaling (IIS) may be critical in diet-induced ADlike pathology. Tau phosphorylation and GSK3 activation mainly result from impaired IIS signaling in the brain [159, 160]. Naryan et al. reported that downregulated IIS increased tau phosphorylation, promoted GSK activation, and decreased insulin receptor substrate-1 (IRS1), phospho-Akt, drebrin, and postsynaptic density (PSD95) resulting in cognitive impairment in the HFD model [156]. Based on these studies, obesity has been identified as one of the major causes for the development of neuropathology, and altered insulin/IGF signaling contributes to obesity-related AD.

Cardiovascular disease-related neuropathology

Hypertension is defined as systolic blood pressure (SBP>140 mmHg) or diastolic blood pressure (DBP>90 mmHg), and is found in more than one billion people worldwide [161]. Hypertension plays a cardinal role in the progression of cerebromicrovascular injury and vascular cognitive impairment [42]. Some studies suggest that changes in cerebral microcirculation play a

Arjunan et al. Cell & Bioscience (2023) 13:16 Page 11 of 18

crucial role in age-related cognitive decline [162-164]. Furthermore, circulating IGF-1 has been shown to be a critical vasoprotective factor that declines with age, and its deficiency can accelerate vascular aging [165]. IGF-1 deficiency is also associated with an increased risk of early atherosclerosis and cerebrovascular disease [165]. Tarantini et al. reported that IGF-1 deficiency accelerates BBB damaged by hypertension, altered capillary morphology in cortical areas, and exacerbates neuroinflammation [42]. Additionally, HFD-fed GH/IGF-1 deficient animals showed glucose intolerance, increased body fat content, oxidative stress, activated inflammatory markers (TNF- α , ICAM-1), and endothelial dysfunction resulting in cerebrovascular damage [166].

Cerebromicrovascular rarefaction leads to decreased cerebral blood flow, which can lead to neurological dysfunction by lowering metabolic factors required for neural signalling [167]. Angelini et al. has been shown that decreased IGF-1 reduced acetylcholine release in the hippocampus, and ultimately led to cognitive decline in hypertensive subjects [43]. Sonntag et al. also showed that IGF-1 influences learning and memory function by regulating K⁺-induced acetylcholine release in the cortex and hippocampus [48]. Endothelium-derived nitric oxide (NO) is a key regulator of microvascular endothelial cell survival and a negative modulator of vascular endothelial growth factor (VEGF) signaling. IGF-1 deficiency impairs endothelial NO bioavailability through elevation of NO breakdown due to increased generation of reactive oxygen species (ROS) and downregulation of endothelial nitric oxide synthase (eNOS) [168, 169].

Other factors contributing to hypertension-related vascular dementia are aging and mitochondrial dysfunction. Reduced mitochondrial biogenesis, neuronal and astrocyte function, and increased ROS are important determinants of aging and neurodegeneration [170–173]. Mitochondria consume about 90% of cellular oxygen through cellular respiration, resulting in a constant stream of free radicals that, if mismanaged, cause long-term oxidative stress and damage [174–176]. IGF-1 reduces the pro-oxidant protein thioredoxin-interacting protein 1 and normalizes ROS levels (Fig. 2). Furthermore, IGF-1 can provide neuroprotection from oxidative danage by interacting with trophic factors secreted by astrocytes in conjunction with $\rm H_2O_2$, such as stem cell factor (SCF) [132].

The IGF-1 pathway is a major determinant of aging. The rate of aging also depends on the amount of IGF-1 and the density of its receptors [177]. There is a considerable increase in neural MAPK phosphorylation with aging along with a decrease in Calcium/Calmodulin-Dependent Protein Kinase IIa (CaMKIIa) levels. Changes in the phosphorylation of synaptic kinases (CaMKII and

MAPK) involved in the regulation of long-term potentiation may be related to IGF-1/IGF-1R signalling [158]. IGF-1 was recently identified as belonging to a new class of ion channel modulators with rapid response (Fig. 2). IGF-1 regulates N-type and L-type Ca²⁺ channels required for neuronal survival and release of neurotransmitters [178]. L-type Ca²⁺ channels (CaV1.2 and CaV1.3) regulate a wide range of neurological functions [179, 180]. IGF-1 can rapidly activate CaV1.3 by modulating IGF-1R, which phosphorylates and activates CaMKII (CaV1.3a and CaV1.3b at the C termini sites, resulting in inositol trisphosphate (IP3)-induced Ca²⁺ release [181]. CaV1.3 phosphorylation by IGF-1 at S1486 residue induces a left-shifted current-voltage that regulates CREB. Excitatory neurotransmitter-induced signaling pathways in the hippocampus are influenced by IGF-1-induced CREB/ CaV1.3 signaling. IGF-1 increased Ca²⁺ influx through L-type Ca²⁺ channels and increased CaMK-IV activity, which reduced the expression of CCAAT enhancer-binding proteins (C/EBPβ) [182].

IGF-1 has been shown to improve mitochondrial function and transmembrane potential in a HFD-fed obese mouse model [183]. IGF-1 contributes considerably to vascular health and protects cells from vascular damage and neuropathological problems [169].

Limitations and future perspectives

Although substantial research has been conducted on the IGF-1 signaling pathway in the past decades, the precise relationship between IGF-1 and cognition remains unclear. Although most studies using animal models have demonstrated neuroprotective effects, human studies have been less conclusive.

Despite its potential therapeutic significance outlined in this review, the long-term benefits of IGF-1 remain controversial. Various side effects have been reported with chronic IGF-1 therapy, including pain at the injection site and lipohypertrophy, headache, hypoglycemia, papilloedema, cataract, neoplasia, renal hypertrophy, and facial nerve palsy [80, 184]. Few studies have shown that overexpression of IGF-1 increases cancer risk through activation of the IRS/Akt/MAPK pathway [31, 80, 81, 120, 185]. Also, Ter Braak et al. mentioned that chronic administration of IGF-1 and its analogue promotes mammary tumor development in the p53R270H/+WAPCre mouse model [186]. Studies have shown that the IGF signaling pathway is not only involved in tumorigenesis, but also contributes to resistance to standard cancer therapies [187]. Whether these unwanted effects may outweigh the benefits in the long run remains an important area of further study. This review has primarily focused on research over the past few decades on the metabolic effects of NDDs. More rigorous studies taking a genetic

GSK3

GRIP

GPF

6-OHDA

VaD

6-Hydroxydopamine

Vascular dementia

Glycogen synthase kinase

Glycine-proline-glutamate

Glutamate receptor AMPAR binding protein

approach are needed to evaluate the role of IGF-1 and its precise downstream mechanistic targets that provide neuroprotection.

Conclusion

GH

Growth hormone

IGF-1 is a master regulator of protein, RNA and DNA synthesis and is involved in Ca²⁺ signaling that regulates synaptogenesis, neurite and glial (astrocytes, oligodendrocytes, schwann cells and microglia) proliferation and repair. Numerous studies have shown the neuroprotective effects of IGF-1. Thus, IGF-1 is a promising therapeutic option for the treatment of various neurological disorders through regulation of multiple neuroprotective signaling pathways, including Ras/Erk1/2, PI3K/MAPK/ Akt/mTOR, Ca2+/CaMK II and IV, CREB, C/EBPB, and GSK3B/NF-kB/NLRP3. It is also involved in regulating neuron and glial homeostasis through regulating ion channels, releasing neurotransmitters, and maintaining synaptic plasticity. Despite significant scientific advances supporting the restorative effects of IGF-1, the precise molecular pathways leading to its neuroprotective effects remain unclear and more studies are needed to accurately understand the role of IGF-1 in MetS-related neurological diseases.

Abbreviations Alzheimer's disease AD Amyloid beta Αβ AMPK AMP-activated protein kinase **AMPAR** α-Amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid receptor Amyloid precursor protein APLP1 Paralogues amyloid precursor-like protein 1 Adenosine triphosphate ATP A AV Adeno-associated viral Bad BCL2 associated agonist of cell death BBB Blood-brain barrier BCI-2 B-Cell leukemia/lymphoma 2 BDNF Brain-derived growth factor BMI Body mass index Calcium ion CaMKII Calcium/calmodulin-dependent protein kinase II CBF Cerebral blood flow CNS Central nervous system CSF Cerebrospinal fluid CP Choroid plexus CI^- Chloride ion CREB CAMP response element binding protein C/EBPB CCAAT enhancer-binding proteins **DAYLs** Disability-adjusted life-years Dopamine DCN Dorsal column nuclei ERK1/2 Extracellular regulated kinase 1/2 DM Diabetes mellitus **eNOS** Endothelial nitric oxide synthase **fEPSPs** Field excitatory postsynaptic potentials FoxO Forkhead box O **GABA** Gamma-aminobutyric acid GBD Global burden of diseases GCL Granule cell layer **GFAP** Glial fibrillary acidic protein

GLUT Glucose transporter GIPC GIPC PDZ domain containing family: member 1 Grb2 Growth factor receptor-bound protein 2 HD Huntington's disease HFD High-fat diet HIF-1 Hypoxia-inducible factor-1 **hNPC** Human neural progenitor cells ICH Intracerebral haemorrhage i.c.v Intracerebroventricular IGF-1 Insulin-like growth factor 1 IGF-1R/2R Insulin-like growth factor 1 receptor Insulin-like growth factor-1/receptor tyrosine kinase IGF-1/RTK **IGFBP** IGF binding protein IIS Insulin/IGF signaling Intraperitoneally i.p iNOS Nitric oxide synthase IP3 Inositol trisphosphate IRS1 Insulin receptor substrate-1 Interleukin Ш JNK C-Jun N-terminal kinases KARs Kainate receptors LID mice Liver IGF-1 deficient mice LID Levodopa-induced dyskinesia I RP1 Low-density lipoprotein receptor-related protein-1 LTP Long-term potentiation MCI Mild cognitive impairment MAPK Mitogen-activated protein kinase MBP Myelin basic protein mtDNA Mitochondrial DNA MT-IGF-1 Monocyte/macrophage-derived IGF-1 NCX Na⁺/Ca²⁺exchanger **NDDs** Neurodegenerative diseases NTF's Neurotrophic factor NMDA N-methyl-p-aspartate Metabolic syndrome MetS MPTP 1-Methyl-4-phenyl-1,2,3,6-tetrahydropyridine MPP+ 1-Methyl-4-phenylpyridine ion NF-ĸB Nuclear factor-κΒ mEPSCs Miniature excitatory postsynaptic currents NSF N-ethylmaleimide-sensitive fusion protein NO Nitric oxide NVC Neurovascular coupling K⁺ Potassium ion PD Parkinson's disease PDK-1 P3 dependent kinase-1 PI3K Phosphatidylinositol 3-kinase PLP Proteolipid protein PSD95 Post synaptic density 95 Ras Rat sarcoma virus RNS Reactive nitrogen species ROS Reactive oxygen species RSKs Ribosomal protein S6 kinase beta SC Schwann cell SCF Stem cell factor sEPSC Spontaneous excitatory postsynaptic current sIPSCs Spontaneous inhibitory postsynaptic currents SNc Substantia nigra compacta Shc Src homology 2 domain containing-transforming protein 1 STZ Streptozotocin SVZ-OB Subventricular zone-olfactory bulb TH Tyrosine hydroxylase TNF-α Tumor necrosis factor-α TIR4 Toll-like receptor 4 T2DM Type 2 diabetes mellitus 7DF Zucker diabetic fatty

Arjunan et al. Cell & Bioscience (2023) 13:16 Page 13 of 18

Acknowledgements

We thank BioRender.com for creating figures.

Author contributions

Writing, AA and JS; Figure, DKS; Manuscript revision, AA and JS; Manuscript finalization, MW and JS. All authors read and approved the final manuscript.

Funding

This study was supported by Grant 2022R1A2C1006125 (Juhyun Song) from the National Research Foundation of Korea (NRF), Republic of Korea. HCRI 22019 from the Chonnam National University Hwasun Hospital Institute for Biomedical Science, Korea (Juhyun Song). The authors acknowledged Biorender.com in creating the figures.

Availability of data and materials

Not applicable.

Declarations

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare no competing interests.

Received: 18 August 2022 Accepted: 17 January 2023 Published online: 23 January 2023

References

- Nilsson PM, Tuomilehto J, Rydén L. The metabolic syndrome—what is it and how should it be managed? Eur J Prev Cardiol. 2019;26(2):33–46.
- Alberti KG, Eckel RH, Grundy SM, Zimmet PZ, Cleeman JI, Donato KA, et al. Harmonizing the metabolic syndrome: a joint interim statement of the International Diabetes Federation Task Force on Epidemiology and Prevention; National Heart, Lung, and Blood Institute; American Heart Association; World Heart Federation; International Atherosclerosis Society; and International Association for the Study of Obesity. Circulation. 2009;120(16):1640–5.
- Van Dyken P, Lacoste B. Impact of metabolic syndrome on neuroinflammation and the blood-brain barrier. Front Neurosci. 2018;12:930.
- Offen D, Shtaif B, Hadad D, Weizman A, Melamed E, Gil-Ad I. Protective effect of insulin-like-growth-factor-1 against dopamine-induced neurotoxicity in human and rodent neuronal cultures: possible implications for Parkinson's disease. Neurosci Lett. 2001;316(3):129–32.
- Niikura T, Hashimoto Y, Okamoto T, Abe Y, Yasukawa T, Kawasumi M, et al. Insulin-like growth factor I (IGF-I) protects cells from apoptosis by Alzheimer's V642I mutant amyloid precursor protein through IGF-I receptor in an IGF-binding protein-sensitive manner. J Neurosci. 2001;21(6):1902–10.
- Herrero-Labrador R, Trueba-Saiz A, Martinez-Rachadell L, de Sevilla MEF, Zegarra-Valdivia JA, Pignatelli J, et al. Circulating insulin-like growth factor I is involved in the effect of high fat diet on peripheral amyloid beta clearance. Int J Mol Sci. 2020. https://doi.org/10.3390/ijms21249675.
- Wang F, Wang L, Wang Y, Li D, Hu T, Sun M, et al. Exogenous IGF-1 improves cognitive function in rats with high-fat diet consumption. J Mol Endocrinol. 2020;64(2):115–23.
- Duarte AI, Petit GH, Ranganathan S, Li JY, Oliveira CR, Brundin P, et al. IGF-1 protects against diabetic features in an in vivo model of Huntington's disease. Exp Neurol. 2011;231(2):314–9.
- 9. Kovacs GG. Molecular pathology of neurodegenerative diseases: principles and practice. J Clin Pathol. 2019;72(11):725–35.
- Dugger BN, Dickson DW. Pathology of neurodegenerative diseases. Cold Spring Harb Perspect Biol. 2017. https://doi.org/10.1101/cshperspect.a028035.

- 11. Gan L, Cookson MR, Petrucelli L, La Spada AR. Converging pathways in neurodegeneration, from genetics to mechanisms. Nat Neurosci. 2018;21(10):1300–9.
- Dharshini SAP, Jemimah S, Taguchi YH, Gromiha MM. Exploring common therapeutic targets for neurodegenerative disorders using transcriptome study. Front Genet. 2021;12: 639160.
- Blesch A. Neurotrophic factors in neurodegeneration. Brain Pathol. 2006;16(4):295–303.
- Costales J, Kolevzon A. The therapeutic potential of insulin-like growth factor-1 in central nervous system disorders. Neurosci Biobehav Rev. 2016;63:207–22.
- 15. Laron Z. Insulin-like growth factor 1 (IGF-1): a growth hormone. Mol Pathol. 2001;54(5):311–6.
- 16. Talia C, Connolly L, Fowler PA. The insulin-like growth factor system: a target for endocrine disruptors? Environ Int. 2021;147: 106311.
- Holly JM, Perks CM. Insulin-like growth factor physiology: what we have learned from human studies. Endocrinol Metab Clin North Am. 2012;41(2):249–63.
- Werner H, LeRoith D. Insulin and insulin-like growth factor receptors in the brain: physiological and pathological aspects. Eur Neuropsychopharmacol. 2014;24(12):1947–53.
- Leung KC, Doyle N, Ballesteros M, Waters MJ, Ho KK. Insulin regulation of human hepatic growth hormone receptors: divergent effects on biosynthesis and surface translocation. J Clin Endocrinol Metab. 2000;85(12):4712–20.
- Frater J, Lie D, Bartlett P, McGrath JJ. Insulin-like Growth Factor 1 (IGF-1) as a marker of cognitive decline in normal ageing: a review. Ageing Res Rev. 2018;42:14–27.
- 21. Bondy CA, Cheng CM. Signaling by insulin-like growth factor 1 in brain. Eur J Pharmacol. 2004;490(1–3):25–31.
- Salzmann A, James SN, Williams DM, Richards M, Cadar D, Schott JM, et al. Investigating the relationship between IGF-I, IGF-II, and IGFBP-3 concentrations and later-life cognition and brain volume. J Clin Endocrinol Metab. 2021;106(6):1617–29.
- 23. Okamoto N, Yoshino K, Kitagawa S, Fujii R, Hamada S, Ikenouchi A, et al. Association between serum insulin-like growth factor 1 levels and the clinical symptoms of chronic schizophrenia: preliminary findings. Front Psychiatry. 2021;12: 653802.
- Carro E, Trejo JL, Gerber A, Loetscher H, Torrado J, Metzger F, et al. Therapeutic actions of insulin-like growth factor I on APP/PS2 mice with severe brain amyloidosis. Neurobiol Aging. 2006;27(9):1250–7.
- Trejo JL, Carro E, Torres-Aleman I. Circulating insulin-like growth factor I mediates exercise-induced increases in the number of new neurons in the adult hippocampus. J Neurosci. 2001;21(5):1628–34.
- 26. Abuzzahab MJ, Schneider A, Goddard A, Grigorescu F, Lautier C, Keller E, et al. IGF-I receptor mutations resulting in intrauterine and postnatal growth retardation. N Engl J Med. 2003;349(23):2211–22.
- Juanes M, Guercio G, Marino R, Berensztein E, Warman DM, Ciaccio M, et al. Three novel IGF1R mutations in microcephalic patients with prenatal and postnatal growth impairment. Clin Endocrinol. 2015;82(5):704–11.
- 28. Park SE, Dantzer R, Kelley KW, McCusker RH. Central administration of insulin-like growth factor-I decreases depressive-like behavior and brain cytokine expression in mice. J Neuroinflammation. 2011;8:12.
- Aghanoori M-R, Agarwal P, Gauvin E, Nagalingam RS, Bonomo R, Yathindranath V, et al. CEBPβ regulation of endogenous IGF-1 in adult sensory neurons can be mobilized to overcome diabetes-induced deficits in bioenergetics and axonal outgrowth. Cell Mol Life Sci. 2022;79(4):1–19.
- Chattopadhyay S, Shubayev VI. MMP-9 controls Schwann cell proliferation and phenotypic remodeling via IGF-1 and ErbB receptor-mediated activation of MEK/ERK pathway. Glia. 2009;57(12):1316–25.
- Cao Y, Gunn AJ, Bennet L, Wu D, George S, Gluckman PD, et al. Insulinlike growth factor (IGF)-1 suppresses oligodendrocyte caspase-3 activation and increases glial proliferation after ischemia in near-term fetal sheep. J Cereb Blood Flow Metab. 2003;23(6):739–47.
- Sun LY, Al-Regaiey K, Masternak MM, Wang J, Bartke A. Local expression of GH and IGF-1 in the hippocampus of GH-deficient long-lived mice. Neurobiol Aging. 2005;26(6):929–37.

- Popken GJ, Hodge RD, Ye P, Zhang J, Ng W, O'Kusky JR, et al. In vivo effects of insulin-like growth factor-I (IGF-I) on prenatal and early postnatal development of the central nervous system. Eur J Neurosci. 2004:19(8):2056–68.
- Ashpole NM, Sanders JE, Hodges EL, Yan H, Sonntag WE. Growth hormone, insulin-like growth factor-1 and the aging brain. Exp Gerontol. 2015;68:76–81.
- 35. Laron Z, Kauli R. Fifty seven years of follow-up of the Israeli cohort of Laron syndrome patients—from discovery to treatment. Growth Horm IGF Res. 2016:28:53–6.
- Lichtenwalner RJ, Forbes ME, Bennett SA, Lynch CD, Sonntag WE, Riddle DR. Intracerebroventricular infusion of insulin-like growth factor-l ameliorates the age-related decline in hippocampal neurogenesis. Neuroscience. 2001;107(4):603–13.
- Åberg MA, Åberg ND, Hedbäcker H, Oscarsson J, Eriksson PS. Peripheral infusion of IGF-I selectively induces neurogenesis in the adult rat hippocampus. J Neurosci. 2000;20(8):2896–903.
- 38. Cheng CM, Cohen M, Tseng V, Bondy CA. Endogenous IGF1 enhances cell survival in the postnatal dentate gyrus. J Neurosci Res. 2001;64(4):341–7.
- Mir S, Cai W, Carlson SW, Saatman KE, Andres DA. IGF-1 mediated neurogenesis involves a novel RIT1/Akt/Sox2 cascade. Sci Rep. 2017;7(1):3283.
- Trejo JL, Piriz J, Llorens-Martin MV, Fernandez AM, Bolós M, LeRoith D, et al. Central actions of liver-derived insulin-like growth factor I underlying its pro-cognitive effects. Mol Psychiatry. 2007;12(12):1118–28.
- Sonntag WE, Ramsey M, Carter CS. Growth hormone and insulin-like growth factor-1 (IGF-1) and their influence on cognitive aging. Ageing Res Rev. 2005;4(2):195–212.
- Tarantini S, Valcarcel-Ares NM, Yabluchanskiy A, Springo Z, Fulop GA, Ashpole N, et al. Insulin-like growth factor 1 deficiency exacerbates hypertension-induced cerebral microhemorrhages in mice, mimicking the aging phenotype. Aging Cell. 2017;16(3):469–79.
- Angelini A, Bendini C, Neviani F, Bergamini L, Manni B, Trenti T, et al. Insulin-like growth factor-1 (IGF-1): relation with cognitive functioning and neuroimaging marker of brain damage in a sample of hypertensive elderly subjects. Arch Gerontol Geriatr. 2009;49(Suppl 1):5–12.
- 44. Fernandez AM, Hernandez-Garzón E, Perez-Domper P, Perez-Alvarez A, Mederos S, Matsui T, et al. Insulin regulates astrocytic glucose handling through cooperation with IGF-I. Diabetes. 2017;66(1):64–74.
- Tarantini S, Balasubramanian P, Yabluchanskiy A, Ashpole NM, Logan S, Kiss T, et al. IGF1R signaling regulates astrocyte-mediated neurovascular coupling in mice: implications for brain aging. GeroScience. 2021:43(2):901–11.
- Calvo D, Gunstad J, Miller LA, Glickman E, Spitznagel MB. Higher serum insulin-like growth factor-1 is associated with better cognitive performance in persons with mild cognitive impairment. Psychogeriatrics. 2013;13(3):170–4.
- Molina DP, Ariwodola OJ, Weiner JL, Brunso-Bechtold JK, Adams MM. Growth hormone and insulin-like growth factor-I alter hippocampal excitatory synaptic transmission in young and old rats. Age. 2013;35(5):1575–87.
- Sonntag WE, Bennett SA, Khan AS, Thornton PL, Xu X, Ingram RL, et al. Age and insulin-like growth factor-1 modulate N-methyl-p-aspartate receptor subtype expression in rats. Brain Res Bull. 2000;51(4):331–8.
- VanGuilder HD, Yan H, Farley JA, Sonntag WE, Freeman WM. Aging alters the expression of neurotransmission-regulating proteins in the hippocampal synaptoproteome. J Neurochem. 2010;113(6):1577–88.
- Kelsch W, Hormuzdi S, Straube E, Lewen A, Monyer H, Misgeld U. Insulin-like growth factor 1 and a cytosolic tyrosine kinase activate chloride outward transport during maturation of hippocampal neurons. J Neurosci. 2001;21(21):8339–47.
- Nuñez A, Carro E, Torres-Aleman I. Insulin-like growth factor I modifies electrophysiological properties of rat brain stem neurons. J Neurophysiol. 2003;89(6):3008–17.
- Aimond F, Rauzier JM, Bony C, Vassort G. Simultaneous activation of p38 MAPK and p42/44 MAPK by ATP stimulates the K+ current ITREK in cardiomyocytes. J Biol Chem. 2000;275(50):39110–6.
- Deijen JB, Arwert LI, Drent ML. The GH/IGF-I axis and cognitive changes across a 4-year period in healthy adults. ISRN Endocrinol. 2011;2011: 249421.

- 54. Yu XW, Oh MM, Disterhoft JF. CREB, cellular excitability, and cognition: implications for aging. Behav Brain Res. 2017;322(Pt B):206–11.
- Zuloaga R, Fuentes EN, Molina A, Valdés JA. The cAMP response element binding protein (CREB) is activated by insulin-like growth factor-1 (IGF-1) and regulates myostatin gene expression in skeletal myoblast. Biochem Biophys Res Commun. 2013;440(2):258–64.
- Merienne K, Pannetier S, Harel-Bellan A, Sassone-Corsi P. Mitogenregulated RSK2-CBP interaction controls their kinase and acetylase activities. Mol Cell Biol. 2001;21(20):7089–96.
- Wiggin GR, Soloaga A, Foster JM, Murray-Tait V, Cohen P, Arthur JS. MSK1 and MSK2 are required for the mitogen- and stress-induced phosphorylation of CREB and ATF1 in fibroblasts. Mol Cell Biol. 2002;22(8):2871–81.
- Qiu C, Kivipelto M, von Strauss E. Epidemiology of Alzheimer's disease: occurrence, determinants, and strategies toward intervention. Dialogues Clin Neurosci. 2009;11(2):111–28.
- Breijyeh Z, Karaman R. Comprehensive review on Alzheimer's disease: causes and treatment. Molecules. 2020. https://doi.org/10.3390/molecules25245789.
- 60. Milionis HJ, Florentin M, Giannopoulos S. Metabolic syndrome and Alzheimer's disease: a link to a vascular hypothesis? CNS Spectr. 2008;13(7):606–13.
- Bye CM, McDonald RJ. A specific role of hippocampal NMDA receptors and Arc protein in rapid encoding of novel environmental representations and a more general long-term consolidation function. Front Behav Neurosci. 2019. https://doi.org/10.3389/fnbeh.2019.00008
- 62. Li F, Tsien JZ. Memory and the NMDA receptors. N Engl J Med. 2009;361(3):302.
- 63. Sumi T, Harada K. Mechanism underlying hippocampal long-term potentiation and depression based on competition between endocytosis and exocytosis of AMPA receptors. Sci Rep. 2020;10(1):14711.
- 64. Bliss TVP, Collingridge GL. Expression of NMDA receptor-dependent LTP in the hippocampus: bridging the divide. Mol Brain. 2013;6(1):5.
- Paoletti P, Bellone C, Zhou Q. NMDA receptor subunit diversity: impact on receptor properties, synaptic plasticity and disease. Nat Rev Neurosci. 2013;14(6):383–400.
- Kakizawa S, Yamada K, Iino M, Watanabe M, Kano M. Effects of insulinlike growth factor I on climbing fibre synapse elimination during cerebellar development. Eur J Neurosci. 2003;17(3):545–54.
- Ramsey MM, Adams MM, Ariwodola OJ, Sonntag WE, Weiner JL. Functional characterization of des-IGF-1 action at excitatory synapses in the CA1 region of rat hippocampus. J Neurophysiol. 2005;94(1):247–54.
- 68. Xing C, Yin Y, Chang R, Gong X, He X, Xie Z. Effects of insulin-like growth factor 1 on synaptic excitability in cultured rat hippocampal neurons. Exp Neurol. 2007;205(1):222–9.
- Shi L, Linville MC, Tucker EW, Sonntag WE, Brunso-Bechtold JK. Differential effects of aging and insulin-like growth factor-1 on synapses in CA1 of rat hippocampus. Cereb Cortex. 2005;15(5):571–7.
- Xing C, Yin Y, Chang R, He X, Xie Z. A role of insulin-like growth factor 1 in beta amyloid-induced disinhibition of hippocampal neurons. Neurosci Lett. 2005;384(1–2):93–7.
- Esteban JA, Shi SH, Wilson C, Nuriya M, Huganir RL, Malinow R. PKA phosphorylation of AMPA receptor subunits controls synaptic trafficking underlying plasticity. Nat Neurosci. 2003;6(2):136–43.
- Bredt DS, Nicoll RA. AMPA receptor trafficking at excitatory synapses. Neuron. 2003;40(2):361–79.
- de la Vega AG, Buño W, Pons S, Garcia-Calderat MS, Garcia-Galloway E, Torres-Aleman I. Insulin-like growth factor I potentiates kainate receptors through a phosphatidylinositol 3-kinase dependent pathway. NeuroReport. 2001;12(6):1293–6.
- Sanchez JC, Lopez-Zapata DF, Francis L, De Los RL. Effects of estradiol and IGF-1 on the sodium calcium exchanger in rat cultured cortical neurons. Cell Mol Neurobiol. 2011;31(4):619–27.
- Xing C, Yin Y, He X, Xie Z. Effects of insulin-like growth factor 1 on voltage-gated ion channels in cultured rat hippocampal neurons. Brain Res. 2006;1072(1):30–5.
- Evans AJ, Gurung S, Henley JM, Nakamura Y, Wilkinson KA. Exciting times: new advances towards understanding the regulation and roles of kainate receptors. Neurochem Res. 2019;44(3):572–84.

 Isaac JTR, Mellor J, Hurtado D, Roche KW. Kainate receptor trafficking: physiological roles and molecular mechanisms. Pharmacol Ther. 2004;104(3):163–72.

(2023) 13:16

- Darstein M, Petralia RS, Swanson GT, Wenthold RJ, Heinemann SF. Distribution of kainate receptor subunits at hippocampal mossy fiber synapses. J Neurosci. 2003;23(22):8013.
- Fadool DA, Tucker K, Phillips JJ, Simmen JA. Brain insulin receptor causes activity-dependent current suppression in the olfactory bulb through multiple phosphorylation of Kv13. J Neurophysiol. 2000;83(4):2332–48.
- Pekic S, Popovic V. Management of endocrine disease: GH therapy and cancer risk in hypopituitarism: what we know from human studies. Eur J Endocrinol. 2013;169(5):R89–97.
- 81. Hua H, Kong Q, Yin J, Zhang J, Jiang Y. Insulin-like growth factor receptor signaling in tumorigenesis and drug resistance: a challenge for cancer therapy. J Hematol Oncol. 2020;13(1):64.
- 82. Westwood AJ, Beiser A, Decarli C, Harris TB, Chen TC, He XM, et al. Insulin-like growth factor-1 and risk of Alzheimer dementia and brain atrophy. Neurology. 2014;82(18):1613–9.
- Carro E, Trejo JL, Gomez-Isla T, LeRoith D, Torres-Aleman I. Serum insulin-like growth factor I regulates brain amyloid-beta levels. Nat Med. 2002;8(12):1390–7.
- Kimoto A, Kasanuki K, Kumagai R, Shibata N, Ichimiya Y, Arai H. Serum insulin-like growth factor-l and amyloid beta protein in Alzheimer's disease: relationship with cognitive function. Psychogeriatrics. 2016;16(4):247–54.
- Kang K, Bai J, Zhong S, Zhang R, Zhang X, Xu Y, et al. Down-regulation of insulin like growth factor 1 involved in Alzheimer's disease via MAPK, Ras, and FoxO signaling pathways. Oxid Med Cell Longev. 2022;2022;8169981.
- Fukudome Y, Tabata T, Miyoshi T, Haruki S, Araishi K, Sawada S, et al. Insulin-like growth factor-I as a promoting factor for cerebellar Purkinje cell development. Eur J Neurosci. 2003;17(10):2006–16.
- 87. Zheng WH, Quirion R. Comparative signaling pathways of insulin-like growth factor-1 and brain-derived neurotrophic factor in hippocampal neurons and the role of the Pl3 kinase pathway in cell survival. J Neurochem. 2004;89(4):844–52.
- 88. Zheng WH, Quirion R. Insulin-like growth factor-1 (IGF-1) induces the activation/phosphorylation of Akt kinase and cAMP response element-binding protein (CREB) by activating different signaling pathways in PC12 cells. BMC Neurosci. 2006;7:51.
- 89. Ma K, Xu H, Zhang J, Zhao F, Liang H, Sun H, et al. Insulin-like growth factor-1 enhances neuroprotective effects of neural stem cell exosomes after spinal cord injury via an miR-219a-2-3p/YY1 mechanism. Aging. 2019;11(24):12278–94.
- Decourt B, Lahiri DK, Sabbagh MN. Targeting tumor necrosis factor alpha for Alzheimer's disease. Curr Alzheimer Res. 2017;14(4):412–25.
- 91. Whiten DR, Brownjohn PW, Moore S, De S, Strano A, Zuo Y, et al. Tumour necrosis factor induces increased production of extracellular amyloid- β -and α -synuclein-containing aggregates by human Alzheimer's disease neurons. Brain Commun. 2020;2(2):fcaa146.
- Johanson CE, Johanson NL. Choroid plexus blood-CSF barrier: major player in brain disease modeling and neuromedicine. J Neurol Neuromed. 2018. https://doi.org/10.29245/2572.942X/2018/4.1194.
- 93. Johanson C, McMillan P, Tavares R, Spangenberger A, Duncan J, Silverberg G, et al. Homeostatic capabilities of the choroid plexus epithelium in Alzheimer's disease. Cerebrospinal Fluid Res. 2004;1(1):3.
- 94. Poewe W, Seppi K, Tanner CM, Halliday GM, Brundin P, Volkmann J, et al. Parkinson disease. Nat Rev Dis Primers. 2017;3:17013.
- 95. Radhakrishnan S, Menon UK, Sundaram KR. Usefulness of a modified questionnaire as a screening tool for swallowing disorders in Parkinson disease: a pilot study. Neurol India. 2019;67(1):118–22.
- Nadjar A, Berton O, Guo S, Leneuve P, Dovero S, Diguet E, et al. IGF-1 signaling reduces neuro-inflammatory response and sensitivity of neurons to MPTP. Neurobiol Aging. 2009;30(12):2021–30.
- Bernhard FP, Heinzel S, Binder G, Weber K, Apel A, Roeben B, et al. Insulin-like growth factor 1 (IGF-1) in Parkinson's disease: potential as trait-, progression- and prediction marker and confounding factors. PLoS ONE. 2016;11(3): e0150552.
- Ghazi Sherbaf F, Mohajer B, Ashraf-Ganjouei A, Mojtahed Zadeh M, Javinani A, Sanjari Moghaddam H, et al. Serum insulin-like growth factor-1 in Parkinson's disease; study of cerebrospinal fluid

- biomarkers and white matter microstructure. Front Endocrinol.
- Godau J, Herfurth M, Kattner B, Gasser T, Berg D. Increased serum insulin-like growth factor 1 in early idiopathic Parkinson's disease. J Neurol Neurosurg Psychiatry. 2010;81(5):536–8.
- Guan J, Krishnamurthi R, Waldvogel HJ, Faull RL, Clark R, Gluckman P. N-terminal tripeptide of IGF-1 (GPE) prevents the loss of TH positive neurons after 6-OHDA induced nigral lesion in rats. Brain Res. 2000;859(2):286–92.
- Krishnamurthi R, Stott S, Maingay M, Faull RL, McCarthy D, Gluckman P, et al. N-terminal tripeptide of IGF-1 improves functional deficits after 6-OHDA lesion in rats. NeuroReport. 2004;15(10):1601–4.
- Quesada A, Micevych PE. Estrogen interacts with the IGF-1 system to protect nigrostriatal dopamine and maintain motoric behavior after 6-hydroxdopamine lesions. J Neurosci Res. 2004;75(1):107–16.
- Ebert AD, Beres AJ, Barber AE, Svendsen CN. Human neural progenitor cells over-expressing IGF-1 protect dopamine neurons and restore function in a rat model of Parkinson's disease. Exp Neurol. 2008;209(1):213–23.
- Pristerà A, Blomeley C, Lopes E, Threlfell S, Merlini E, Burdakov D, et al. Dopamine neuron-derived IGF-1 controls dopamine neuron firing, skill learning, and exploration. Proc Natl Acad Sci USA. 2019;116(9):3817–26.
- Quesada A, Lee BY, Micevych PE. PI3 kinase/Akt activation mediates estrogen and IGF-1 nigral DA neuronal neuroprotection against a unilateral rat model of Parkinson's disease. Dev Neurobiol. 2008;68(5):632–44.
- 106. Wang L, Yang HJ, Xia YY, Feng ZW. Insulin-like growth factor 1 protects human neuroblastoma cells SH-EP1 against MPP+induced apoptosis by AKT/GSK-3β/JNK signaling. Apoptosis. 2010;15(12):1470–9.
- 107. Alberti KG, Zimmet P, Shaw J. The metabolic syndrome—a new worldwide definition. Lancet. 2005;366(9491):1059–62.
- Saklayen MG. The global epidemic of the metabolic syndrome. Curr Hypertens Rep. 2018;20(2):12.
- Watts AS, Loskutova N, Burns JM, Johnson DK. Metabolic syndrome and cognitive decline in early Alzheimer's disease and healthy older adults. J Alzheimer's Dis. 2013;35(2):253–65.
- Wang F, Zhao M, Han Z, Li D, Zhang S, Zhang Y, et al. Long-term subclinical hyperglycemia and hypoglycemia as independent risk factors for mild cognitive impairment in elderly people. Tohoku J Exp Med. 2017;242(2):121–8.
- 111. Zhong Y, Zhu Y, He T, Li W, Li Q, Miao Y. Brain-derived neurotrophic factor inhibits hyperglycemia-induced apoptosis and downregulation of synaptic plasticity-related proteins in hippocampal neurons via the PI3K/Akt pathway. Int J Mol Med. 2019;43(1):294–304.
- Mukherjee A, Mehta BK, Sen KK, Banerjee S. Metabolic syndromeassociated cognitive decline in mice: role of minocycline. Indian J Pharmacol. 2018;50(2):61–8.
- Galicia-Garcia U, Benito-Vicente A, Jebari S, Larrea-Sebal A, Siddiqi H, Uribe KB, et al. Pathophysiology of type 2 diabetes mellitus. Int J Mol Sci. 2020. https://doi.org/10.3390/ijms21176275.
- 114. Robertson K, Lu Y, De Jesus K, Li B, Su Q, Lund PK, et al. A general and islet cell-enriched overexpression of IGF-I results in normal islet cell growth, hypoglycemia, and significant resistance to experimental diabetes. Am J Physiol Endocrinol Metab. 2008;294(5):E928–38.
- Spauwen PJ, Köhler S, Verhey FR, Stehouwer CD, van Boxtel MP. Effects of type 2 diabetes on 12-year cognitive change: results from the Maastricht aging study. Diabetes Care. 2013;36(6):1554–61.
- Li X, Song D, Leng SX. Link between type 2 diabetes and Alzheimer's disease: from epidemiology to mechanism and treatment. Clin Interv Aging. 2015;10:549–60.
- Mittal K, Katare DP. Shared links between type 2 diabetes mellitus and Alzheimer's disease: a review. Diabetes Metab Syndr. 2016;10(2 Suppl 1):S144-9.
- Adzovic L, Lynn AE, D'Angelo HM, Crockett AM, Kaercher RM, Royer SE, et al. Insulin improves memory and reduces chronic neuroinflammation in the hippocampus of young but not aged brains. J Neuroinflammation. 2015;12:63.

- 119. Benedict C, Hallschmid M, Hatke A, Schultes B, Fehm HL, Born J, et al. Intranasal insulin improves memory in humans. Psychoneuroendocrinology. 2004;29(10):1326–34.
- 120. Ortiz GG, Huerta M, Gonzalez-Usigli HA, Torres-Sanchez ED, Delgado-Lara DL, Pacheco-Moises FP, et al. Cognitive disorder and dementia in type 2 diabetes mellitus. World J Diabetes. 2022;13(4):319–37.
- Yuan XY, Wang XG. Mild cognitive impairment in type 2 diabetes mellitus and related risk factors: a review. Rev Neurosci. 2017;28(7):715–23.
- 122. Rui-Hua C, Yong-de P, Xiao-Zhen J, Chen J, Bin Z. Decreased levels of serum IGF-1 and vitamin D are associated with cognitive impairment in patients with type 2 diabetes. Am J Alzheimers Dis Other Demen. 2019;34(7–8):450–6.
- 123. Huang R, Wang P, Han J, Xia W, Cai R, Sun H, et al. Decreased serum IGF-1/IGFBP-3 Molar ratio is associated with executive function behaviors in type 2 diabetic patients with mild cognitive impairment. J Alzheimer's Dis. 2015;48(3):875.
- Aksu I, Ates M, Baykara B, Kiray M, Sisman AR, Buyuk E, et al. Anxiety correlates to decreased blood and prefrontal cortex IGF-1 levels in streptozotocin induced diabetes. Neurosci Lett. 2012;531(2):176–81.
- 125. Jing YH, Song YF, Yao YM, Yin J, Wang DG, Gao LP. Retardation of fetal dendritic development induced by gestational hyperglycemia is associated with brain insulin/IGF-I signals. Int J Dev Neurosci. 2014;37:15–20.
- Srinivasan S, Stevens M, Wiley JW. Diabetic peripheral neuropathy: evidence for apoptosis and associated mitochondrial dysfunction. Diabetes. 2000;49(11):1932–8.
- Vinik Al, Casellini CM. Guidelines in the management of diabetic nerve pain: clinical utility of pregabalin. Diabetes Metab Syndr Obes. 2013;6:57–78.
- Chu Q, Moreland R, Yew NS, Foley J, Ziegler R, Scheule RK. Systemic Insulin-like growth factor-1 reverses hypoalgesia and improves mobility in a mouse model of diabetic peripheral neuropathy. Mol Ther. 2008;16(8):1400–8.
- Russell JW, Cheng H-L, Golovoy D. Insulin-like growth factor-i promotes myelination of peripheral sensory axons. J Neuropathol Exp Neurol. 2000;59(7):575–84.
- 130. Ye P, Li L, Lund PK, D'Ercole AJ. Deficient expression of insulin receptor substrate-1 (IRS-1) fails to block insulin-like growth factor-I (IGF-I) stimulation of brain growth and myelination. Brain Res Dev Brain Res. 2002;136(2):111–21.
- Dávila D, Fernández S, Torres-Alemán I. Astrocyte resilience to oxidative stress induced by insulin-like growth factor I (IGF-I) involves preserved AKT (protein kinase B) activity. J Biol Chem. 2016;291(23):12039.
- 132. Genis L, Dávila D, Fernandez S, Pozo-Rodrigálvarez A, Martínez-Murillo R, Torres-Aleman I. Astrocytes require insulin-like growth factor I to protect neurons against oxidative injury. F1000Res. 2014;3:28.
- 133. Yu J, Li J, Zhang S, Xu X, Zheng M, Jiang G, et al. IGF-1 induces hypoxiainducible factor 1α-mediated GLUT3 expression through PI3K/Akt/ mTOR dependent pathways in PC12 cells. Brain Res. 2012;1430:18–24.
- 134. Chen X, Le Y, Tang S-Q, He W-y, He J, Wang Y-h, et al. Painful Diabetic Neuropathy Is Associated with Compromised Microglial IGF-1 Signaling Which Can Be Rescued by Green Tea Polyphenol EGCG in Mice. Oxidative medicine and cellular longevity. 2022;2022:6773662.
- Ransohoff RM. A polarizing question: do M1 and M2 microglia exist? Nat Neurosci. 2016;19(8):987–91.
- Kettenmann H, Hanisch UK, Noda M, Verkhratsky A. Physiology of microglia. Physiol Rev. 2011;91(2):461–553.
- Labandeira-Garcia JL, Costa-Besada MA, Labandeira CM, Villar-Cheda B, Rodríguez-Perez Al. Insulin-like growth factor-1 and neuroinflammation. Front Aging Neurosci. 2017. https://doi.org/10.3389/fnagi.2017.00365.
- Labandeira-Garcia JL, Rodríguez-Perez AI, Garrido-Gil P, Rodriguez-Pallares J, Lanciego JL, Guerra MJ. Brain renin-angiotensin system and microglial polarization: implications for aging and neurodegeneration. Front Aging Neurosci. 2017. https://doi.org/10.3389/fnagi.2017.0012.
- 139. Suh HS, Zhao ML, Derico L, Choi N, Lee SC. Insulin-like growth factor 1 and 2 (IGF1, IGF2) expression in human microglia: differential regulation by inflammatory mediators. J Neuroinflammation. 2013;10:37.
- Spadaro O, Camell CD, Bosurgi L, Nguyen KY, Youm YH, Rothlin CV, et al. IGF1 shapes macrophage activation in response to immunometabolic challenge. Cell Rep. 2017;19(2):225–34.

- 141. Sun Z, Wu K, Gu L, Huang L, Zhuge Q, Yang S, et al. IGF-1R stimulation alters microglial polarization via TLR4/NF-kappaB pathway after cerebral hemorrhage in mice. Brain Res Bull. 2020;164:221–34.
- 142. Mirdamadī Y, Bommhardt U, Goihl A, Guttek K, Zouboulis CC, Quist S, et al. Insulin and Insulin-like growth factor-1 can activate the phosphoinositide-3-kinase/Akt/FoxO1 pathway in T cells in vitro. Dermatoendocrinol. 2017;9(1): e1356518.
- Obadia N, Andrade G, Leardini-Tristao M, Albuquerque L, Garcia C, Lima F, et al. TLR4 mutation protects neurovascular function and cognitive decline in high-fat diet-fed mice. J Neuroinflammation. 2022;19(1):104.
- 144. Bellini MJ, Hereñú CB, Goya RG, Garcia-Segura LM. Insulin-like growth factor-I gene delivery to astrocytes reduces their inflammatory response to lipopolysaccharide. J Neuroinflammation. 2011;8(1):21.
- Wolters TLC, Netea MG, Hermus A, Smit JWA, Netea-Maier RT. IGF1
 potentiates the pro-inflammatory response in human peripheral blood
 mononuclear cells via MAPK. J Mol Endocrinol. 2017;59(2):129–39.
- Lee WJ. IGF-I exerts an anti-inflammatory effect on skeletal muscle cells through down-regulation of TLR4 signaling. Immune Netw. 2011;11(4):223–6.
- Finicelli M, Squillaro T, Di Cristo F, Di Salle A, Melone MAB, Galderisi U, et al. Metabolic syndrome, Mediterranean diet, and polyphenols: evidence and perspectives. J Cell Physiol. 2019;234(5):5807–26.
- 148. Ng M, Fleming T, Robinson M, Thomson B, Graetz N, Margono C, et al. Global, regional, and national prevalence of overweight and obesity in children and adults during 1980–2013: a systematic analysis for the Global Burden of Disease Study 2013. Lancet. 2014;384(9945):766–81.
- Misra A, Khurana L. Obesity and the metabolic syndrome in developing countries. J Clin Endocrinol Metab. 2008;93(11 Suppl 1):S9-30.
- Deckers K, Van Boxtel MPJ, Verhey FRJ, Köhler S. Obesity and cognitive decline in adults: effect of methodological choices and confounding by age in a longitudinal study. J Nutr Health Aging. 2017;21(5):546–53.
- Ganguli M, Beer JC, Zmuda JM, Ryan CM, Sullivan KJ, Chang CH, et al. Aging, diabetes, obesity, and cognitive decline: a population-based study. J Am Geriatr Soc. 2020;68(5):991–8.
- Chen J, Guan Z, Wang L, Song G, Ma B, Wang Y. Meta-analysis: overweight, obesity, and Parkinson's disease. Int J Endocrinol. 2014;2014: 203930
- Palacios N, Gao X, McCullough ML, Jacobs EJ, Patel AV, Mayo T, et al. Obesity, diabetes, and risk of Parkinson's disease. Mov Disord. 2011;26(12):2253–9.
- Waldstein SR, Katzel LI. Interactive relations of central versus total obesity and blood pressure to cognitive function. Int J Obes. 2006;30(1):201–7.
- Wang GJ, Volkow ND, Logan J, Pappas NR, Wong CT, Zhu W, et al. Brain dopamine and obesity. Lancet. 2001;357(9253):354–7.
- Bhat NR, Thirumangalakudi L. Increased tau phosphorylation and impaired brain insulin/IGF signaling in mice fed a high fat/high cholesterol diet. J Alzheimer's Dis. 2013;36(4):781–9.
- 157. Guerra-Cantera S, Frago LM, Jiménez-Hernaiz M, Ros P, Freire-Regatillo A, Barrios V, et al. Impact of long-term HFD intake on the peripheral and central IGF system in male and female mice. Metabolites. 2020;10(11):462.
- 158. Ogundele OM, Pardo J, Francis J, Goya RG, Lee CC. A putative mechanism of age-related synaptic dysfunction based on the impact of IGF-1 receptor signaling on synaptic CaMKIIa phosphorylation. Front Neuroanat. 2018. https://doi.org/10.3389/fnana.2018.00035.
- Sun MK, Alkon DL. Links between Alzheimer's disease and diabetes. Drugs of today. 2006;42(7):481–9.
- Pasinetti GM, Eberstein JA. Metabolic syndrome and the role of dietary lifestyles in Alzheimer's disease. J Neurochem. 2008;106(4):1503–14.
- 161. James PA, Oparil S, Carter BL, Cushman WC, Dennison-Himmelfarb C, Handler J, et al. 2014 evidence-based guideline for the management of high blood pressure in adults: report from the panel members appointed to the Eighth Joint National Committee (JNC 8). JAMA. 2014;311(5):507–20.
- Canavan M, O'Donnell MJ. Hypertension and cognitive impairment: a review of mechanisms and key concepts. Front Neurol. 2022;13: 821135.
- Toth P, Tucsek Z, Sosnowska D, Gautam T, Mitschelen M, Tarantini S, et al. Age-related autoregulatory dysfunction and cerebromicrovascular

- injury in mice with angiotensin Il-induced hypertension. J Cereb Blood Flow Metab. 2013;33(11):1732–42.
- 164. Toth P, Tucsek Z, Tarantini S, Sosnowska D, Gautam T, Mitschelen M, et al. IGF-1 deficiency impairs cerebral myogenic autoregulation in hypertensive mice. J Cereb Blood Flow Metab. 2014;34(12):1887–97.
- Ungvari Z, Csiszar A. The emerging role of IGF-1 deficiency in cardiovascular aging: recent advances. J Gerontol A Biol Sci Med Sci. 2012;67(6):599–610.
- 166. Bailey-Downs LC, Sosnowska D, Toth P, Mitschelen M, Gautam T, Henthorn JC, et al. Growth hormone and IGF-1 deficiency exacerbate high-fat diet-induced endothelial impairment in obese Lewis dwarf rats: implications for vascular aging. J Gerontol A Biol Sci Med Sci. 2012;67(6):553–64.
- 167. Riddle DR, Sonntag WE, Lichtenwalner RJ. Microvascular plasticity in aging. Ageing Res Rev. 2003;2(2):149–68.
- Csiszar A, Labinskyy N, Perez V, Recchia FA, Podlutsky A, Mukhopadhyay P, et al. Endothelial function and vascular oxidative stress in long-lived GH/IGF-deficient Ames dwarf mice. Am J Physiol Heart Circ Physiol. 2008:295(5):H1882–94.
- 169. Toth P, Tarantini S, Ashpole NM, Tucsek Z, Milne GL, Valcarcel-Ares NM, et al. IGF-1 deficiency impairs neurovascular coupling in mice: implications for cerebromicrovascular aging. Aging Cell. 2015;14(6):1034–44.
- 170. Kubik LL, Philbert MA. The role of astrocyte mitochondria in differential regional susceptibility to environmental neurotoxicants: tools for understanding neurodegeneration. Toxicol Sci. 2015;144(1):7–16.
- 171. Parihar MS, Kunz EA, Brewer GJ. Age-related decreases in NAD(P) H and glutathione cause redox declines before ATP loss during glutamate treatment of hippocampal neurons. J Neurosci Res. 2008;86(10):2339–52.
- 172. Parihar MS, Brewer GJ. Mitoenergetic failure in Alzheimer disease. Am J Physiol Cell Physiol. 2007;292(1):C8-23.
- 173. Vančová O, Bačiak L, Kašparová S, Kucharská J, Palacios HH, Horecký J, et al. In vivo and in vitro assessment of brain bioenergetics in aging rats. J Cell Mol Med. 2010;14(11):2667–74.
- 174. Elfawy HA, Das B. Crosstalk between mitochondrial dysfunction, oxidative stress, and age related neurodegenerative disease: etiologies and therapeutic strategies. Life Sci. 2019;218:165–84.
- 175. Guo C, Sun L, Chen X, Zhang D. Oxidative stress, mitochondrial damage and neurodegenerative diseases. Neural Regen Res. 2013;8(21):2003–14.
- 176. Islam MT. Oxidative stress and mitochondrial dysfunction-linked neurodegenerative disorders. Neurol Res. 2017;39(1):73–82.
- 177. Bartke A, List EO, Kopchick JJ. The somatotropic axis and aging: benefits of endocrine defects. Growth Horm IGF Res. 2016;27:41–5.
- Ge L, Liu S, Rubin L, Lazarovici P, Zheng W. Research progress on neuroprotection of insulin-like growth factor-1 towards glutamate-induced neurotoxicity. Cells. 2022. https://doi.org/10.3390/cells11040666.
- Redmond L, Kashani AH, Ghosh A. Calcium regulation of dendritic growth via CaM kinase IV and CREB-mediated transcription. Neuron. 2002;34(6):999–1010.
- Yamamoto K, Sakagami Y, Sugiura S, Inokuchi K, Shimohama S, Kato N. Homer 1a enhances spike-induced calcium influx via L-type calcium channels in neocortex pyramidal cells. Eur J Neurosci. 2005;22(6):1338–48.
- 181. Gao L, Blair LAC, Salinas GD, Needleman LA, Marshall J. Insulin-like growth factor-1 modulation of CaV1.3 calcium channels depends on Ca2+ release from IP3-sensitive stores and calcium/calmodulin kinase II phosphorylation of the alpha1 subunit EF hand. J Neurosci. 2006;26(23):6259–68.
- 182. Marshall J, Dolan BM, Garcia EP, Sathe S, Tang X, Mao Z, et al. Calcium channel and NMDA receptor activities differentially regulate nuclear C/ EBPbeta levels to control neuronal survival. Neuron. 2003;39(4):625–39.
- Yang C, Sui G, Li D, Wang L, Zhang S, Lei P, et al. Exogenous IGF-1 alleviates depression-like behavior and hippocampal mitochondrial dysfunction in high-fat diet mice. Physiol Behav. 2021;229: 113236.
- 184. Backeljauw PF, Underwood LE. Therapy for 6.5–7.5 years with recombinant insulin-like growth factor I in children with growth hormone insensitivity syndrome: a clinical research center study. J Clin Endocrinol Metab. 2001;86(4):1504–10.
- 185. Torres-Aleman I. Toward a comprehensive neurobiology of IGF-I. Dev Neurobiol. 2010;70(5):384–96.

- 186. ter Braak B, Siezen C, Speksnijder EN, Koedoot E, van Steeg H, Salvatori DC, et al. Mammary gland tumor promotion by chronic administration of IGF1 and the insulin analogue AspB10 in the p53R270H/(+)WAPCre mouse model. Breast Cancer Res. 2015;17:14.
- 187. Heskamp S, Boerman OC, Molkenboer-Kuenen JD, Wauters CA, Strobbe LJ, Mandigers CM, et al. Upregulation of IGF-1R expression during neoadjuvant therapy predicts poor outcome in breast cancer patients. PLoS ONE. 2015;10(2): e0117745.
- 188. Hayes CA, Valcarcel-Ares MN, Ashpole NM. Preclinical and clinical evidence of IGF-1 as a prognostic marker and acute intervention with ischemic stroke. J Cereb Blood Flow Metab. 2021;41(10):2475–91.
- 189. Tarantini S, Nyúl-Tóth Á, Yabluchanskiy A, Csipo T, Mukli P, Balasubramanian P, et al. Endothelial deficiency of insulin-like growth factor-1 receptor (IGF1R) impairs neurovascular coupling responses in mice, mimicking aspects of the brain aging phenotype. GeroScience. 2021;43(5):2387–94.
- 190. Horvath A, Salman Z, Quinlan P, Wallin A, Svensson J. Patients with Alzheimer's disease have increased levels of insulin-like growth factor-i in serum but not in cerebrospinal fluid. J Alzheimer's Dis. 2020;75(1):289–98.
- Pharaoh G, Owen D, Yeganeh A, Premkumar P, Farley J, Bhaskaran S, et al. Disparate central and peripheral effects of circulating IGF-1 deficiency on tissue mitochondrial function. Mol Neurobiol. 2020;57(3):1317–31.
- Prabhu D, Khan SM, Blackburn K, Marshall JP, Ashpole NM. Loss of insulin-like growth factor-1 signaling in astrocytes disrupts glutamate handling. J Neurochem. 2019;151(6):689–702.
- 193. Logan S, Pharaoh GA, Marlin MC, Masser DR, Matsuzaki S, Wronowski B, et al. Insulin-like growth factor receptor signaling regulates working memory, mitochondrial metabolism, and amyloid-β uptake in astrocytes. Mol Metab. 2018;9:141–55.
- 194. Nageeb RS, Hashim NA, Fawzy A. Serum insulin-like growth factor 1 (IGF-1) in multiple sclerosis: relation to cognitive impairment and fatigue. Egypt J Neurol Psychiatry Neurosurg. 2018;54(1):25.
- Quinlan P, Horvath A, Nordlund A, Wallin A, Svensson J. Low serum insulin-like growth factor-I (IGF-I) level is associated with increased risk of vascular dementia. Psychoneuroendocrinology. 2017;86:169–75.
- 196. Vidal JS, Hanon O, Funalot B, Brunel N, Viollet C, Rigaud AS, et al. Low serum insulin-like growth factor-i predicts cognitive decline in Alzheimer's disease. J Alzheimer's Dis. 2016;52(2):641–9.
- 197. Hu X, Yang Y, Gong D. Circulating insulin-like growth factor 1 and insulin-like growth factor binding protein-3 level in Alzheimer's disease: a meta-analysis. Neurol Sci. 2016;37(10):1671–7.
- 198. Nieto-Estevez V, Oueslati-Morales CO, Li L, Pickel J, Morales AV, Vicario-Abejon C. Brain insulin-like growth factor-i directs the transition from stem cells to mature neurons during postnatal/adult hippocampal neurogenesis. Stem Cells. 2016;34(8):2194–209.
- 199. Basta-Kaim A, Szczesny E, Glombik K, Stachowicz K, Slusarczyk J, Nalepa I, et al. Prenatal stress affects insulin-like growth factor-1 (IGF-1) level and IGF-1 receptor phosphorylation in the brain of adult rats. Eur Neuropsychopharmacol. 2014;24(9):1546–56.
- Johansson P, Åberg D, Johansson J-O, Mattsson N, Hansson O, Ahrén B, et al. Serum but not cerebrospinal fluid levels of insulin-like growth factor-I (IGF-I) and IGF-binding protein-3 (IGFBP-3) are increased in Alzheimer's disease. Psychoneuroendocrinology. 2013;38(9):1729–37.
- Duron E, Funalot B, Brunel N, Coste J, Quinquis L, Viollet C, et al. Insulinlike growth factor-I and insulin-like growth factor binding protein-3 in Alzheimer's disease. J Clin Endocrinol Metab. 2012;97(12):4673–81.
- Mitschelen M, Yan H, Farley JA, Warrington JP, Han S, Hereñú CB, et al. Long-term deficiency of circulating and hippocampal insulin-like growth factor I induces depressive behavior in adult mice: a potential model of geriatric depression. Neuroscience. 2011;185:50–60.
- Moloney AM, Griffin RJ, Timmons S, O'Connor R, Ravid R, O'Neill C. Defects in IGF-1 receptor, insulin receptor and IRS-1/2 in Alzheimer's disease indicate possible resistance to IGF-1 and insulin signalling. Neurobiol Aging. 2010;31(2):224–43.
- Freude S, Hettich MM, Schumann C, Stöhr O, Koch L, Köhler C, et al. Neuronal IGF-1 resistance reduces Aβ accumulation and protects against premature death in a model of Alzheimer's disease. FASEB J. 2009;23(10):3315–24.

Arjunan et al. Cell & Bioscience (2023) 13:16 Page 18 of 18

- Watanabe T, Miyazaki A, Katagiri T, Yamamoto H, Idei T, Iguchi T. Relationship between serum insulin-like growth factor-1 levels and Alzheimer's disease and vascular dementia. J Am Geriatr Soc. 2005;53(10):1748–53.
- 206. Selles MC, Fortuna JTS, Zappa-Villar MF, de Faria YPR, Souza AS, Suemoto CK, et al. Adenovirus-mediated transduction of insulin-like growth factor 1 protects hippocampal neurons from the toxicity of abeta oligomers and prevents memory loss in an Alzheimer mouse model. Mol Neurobiol. 2020;57(3):1473–83.
- Farias Quipildor GE, Mao K, Hu Z, Novaj A, Cui M-H, Gulinello M, et al. Central IGF-1 protects against features of cognitive and sensorimotor decline with aging in male mice. GeroScience. 2019;41(2):185–208.
- Carlson SW, Saatman KE. Central infusion of insulin-like growth factor-1 increases hippocampal neurogenesis and improves neurobehavioral function after traumatic brain injury. J Neurotrauma. 2018;35(13):1467–80.
- 209. Morel GR, Leon ML, Uriarte M, Reggiani PC, Goya RG. Therapeutic potential of IGF-I on hippocampal neurogenesis and function during aging. Neurogenesis. 2017;4(1): e1259709.
- Pardo J, Uriarte M, Cónsole GM, Reggiani PC, Outeiro TF, Morel GR, et al. Insulin-like growth factor-I gene therapy increases hippocampal neurogenesis, astrocyte branching and improves spatial memory in female aging rats. Eur J Neurosci. 2016;44(4):2120–8.
- Bake S, Selvamani A, Cherry J, Sohrabji F. Blood brain barrier and neuroinflammation are critical targets of IGF-1-mediated neuroprotection in stroke for middle-aged female rats. PLoS ONE. 2014;9(3):e91427.
- Jacobsen KT, Adlerz L, Multhaup G, Iverfeldt K. Insulin-like growth factor-1 (IGF-1)-induced Processing of amyloid-β precursor protein (APP) and APP-like protein 2 is mediated by different metalloproteinases. J Biol Chem. 2010;285(14):10223–31.
- 213. Sun X, Huang L, Zhang M, Sun S, Wu Y. Insulin like growth factor-1 prevents 1-mentyl-4-phenylphyridinium-induced apoptosis in PC12 cells through activation of glycogen synthase kinase-3beta. Toxicology. 2010:271(1–2):5–12.
- Kao S-Y. Rescue of a-synuclein cytotoxicity by insulin-like growth factors. Biochem Biophys Res Commun. 2009;385(3):434–8.
- 215. Aghanoori M-R, Smith DR, Shariati-levari S, Ajisebutu A, Nguyen A, Desmond F, et al. Insulin-like growth factor-1 activates AMPK to augment mitochondrial function and correct neuronal metabolism in sensory neurons in type 1 diabetes. Mol Metab. 2019;20:149–65.
- 216. Rizk NN, Myatt-Jones J, Rafols J, Dunbar JC. Insulin like growth factor-1 (IGF-1) decreases ischemia-reperfusion induced apoptosis and necrosis in diabetic rats. Endocrine. 2007;31(1):66–71.
- Wold LE, Muralikrishnan D, Albano CB, Norby FL, Ebadi M, Ren J. Insulinlike growth factor I (IGF-1) supplementation prevents diabetes-induced alterations in coenzymes Q9 and Q10. Acta Diabetol. 2003;40(2):85–90.
- 218. Delaney CL, Russell JW, Cheng HL, Feldman EL. Insulin-like growth factor-I and over-expression of Bcl-xL prevent glucose-mediated apoptosis in Schwann cells. J Neuropathol Exp Neurol. 2001;60(2):147–60.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- $\bullet\,$ thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

